

Movies of proteins at work

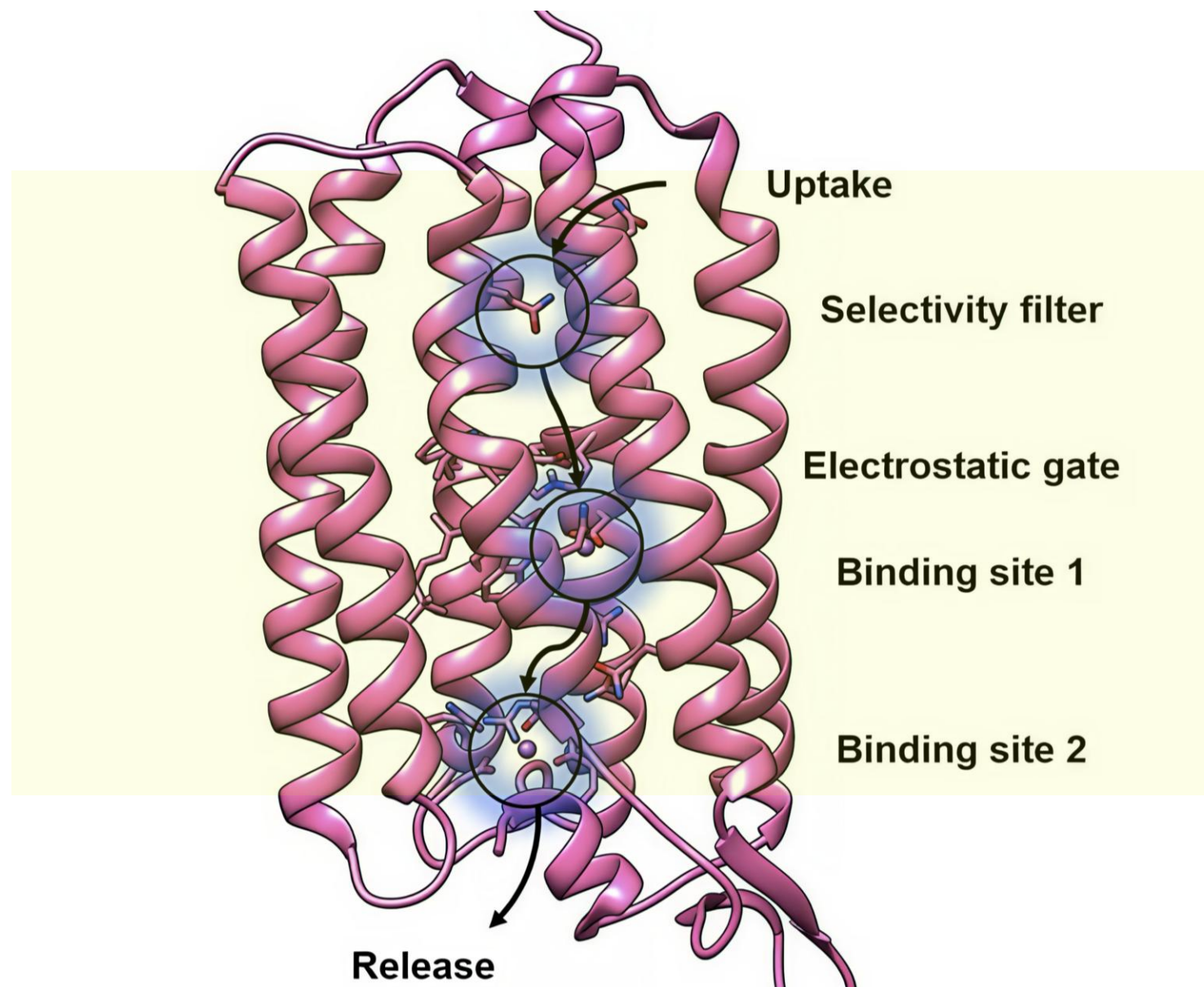
— and why they're harder to make than they look

Tobias Weinert · Senior Scientist · Paul Scherrer Institute · PSI Center for Life Sciences

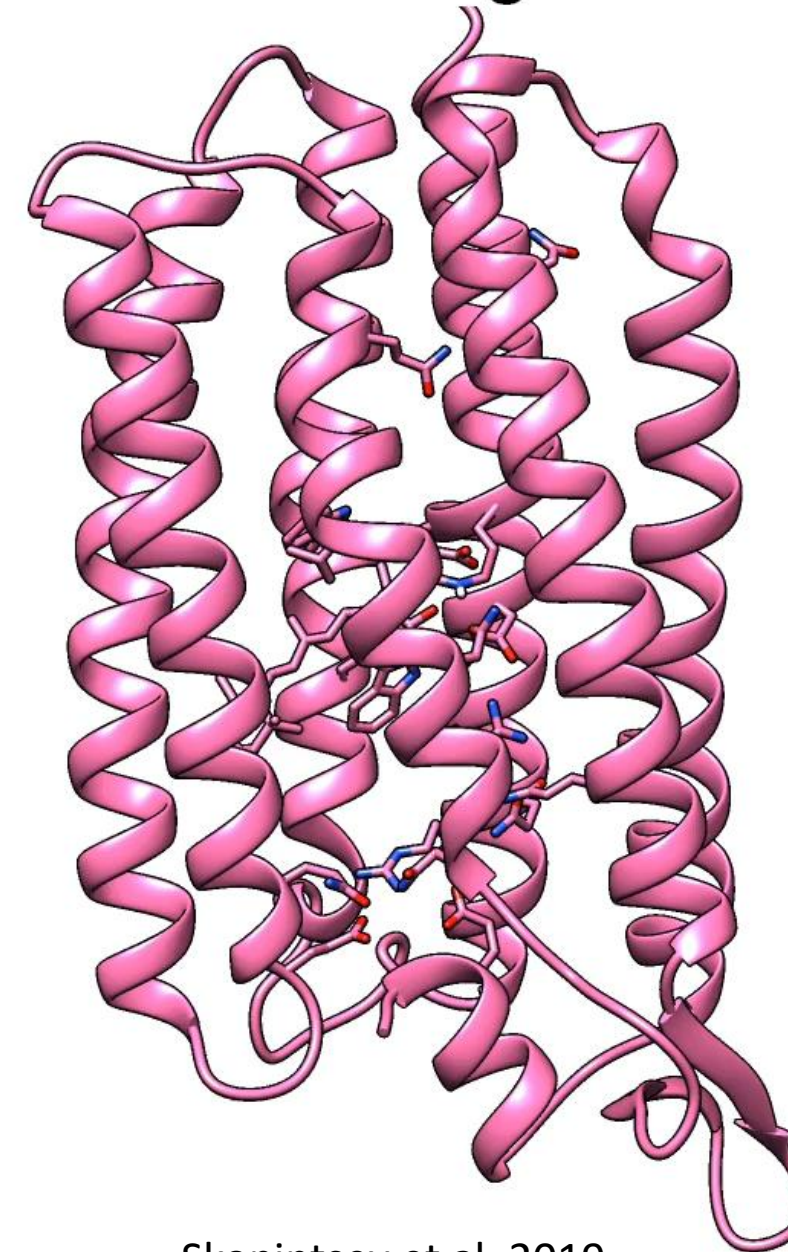
VII ALBA Users' Meeting · 04.09.2026

Why movies?

A three-dimensional movie can explain function directly



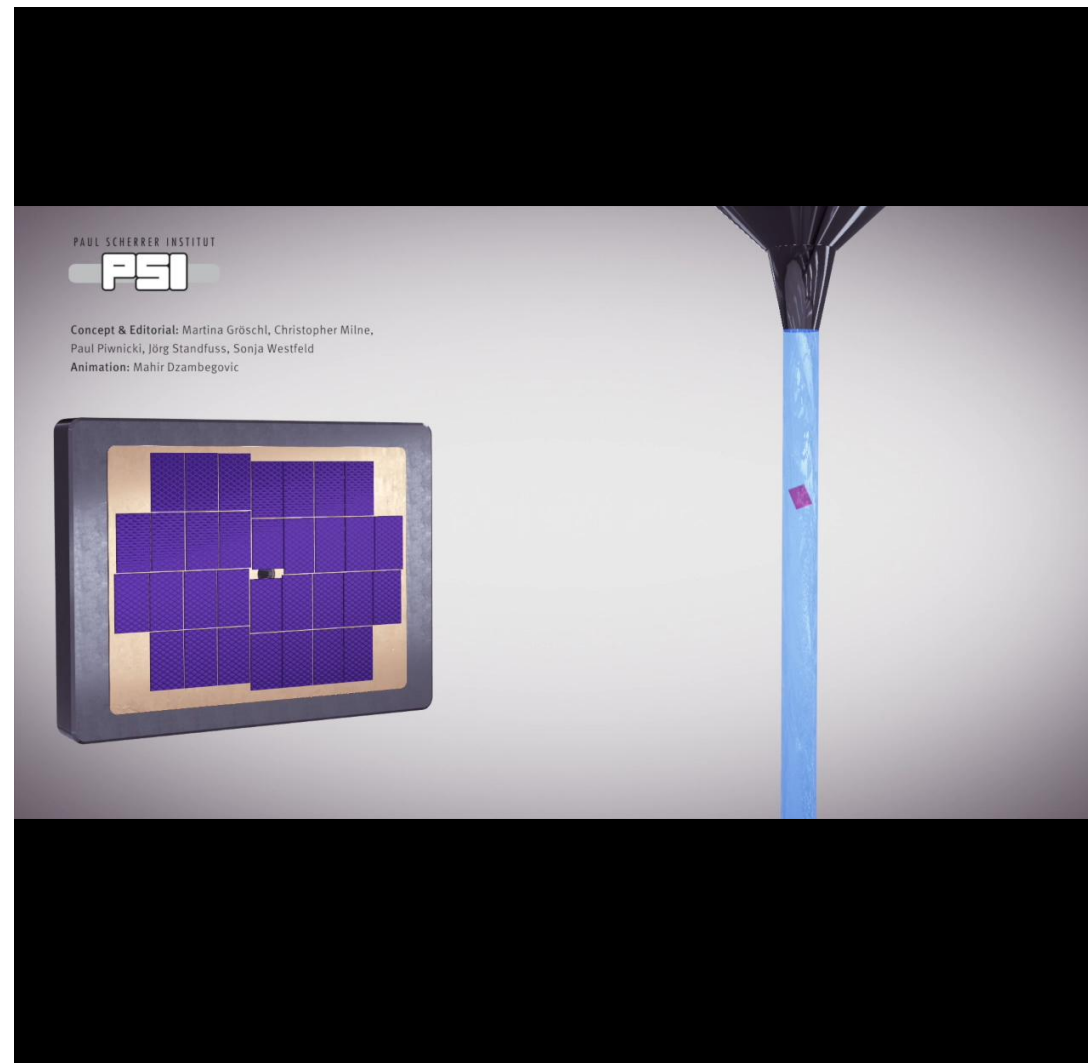
KR2 resting state



Skopintsev et al. 2019

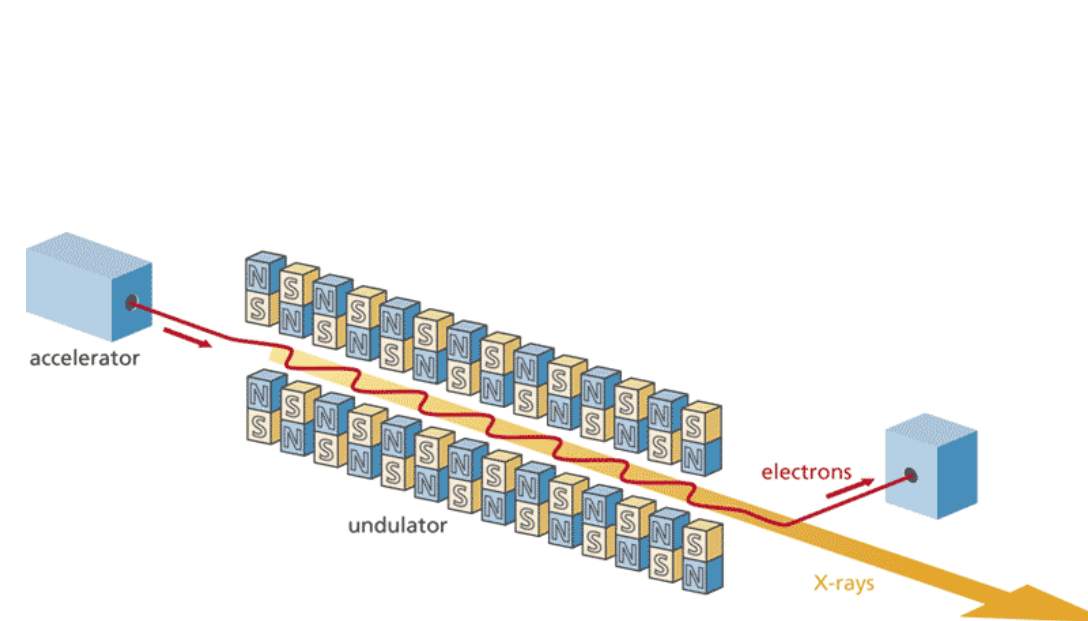
How can we make them?

Three things you need for a time-resolved experiment



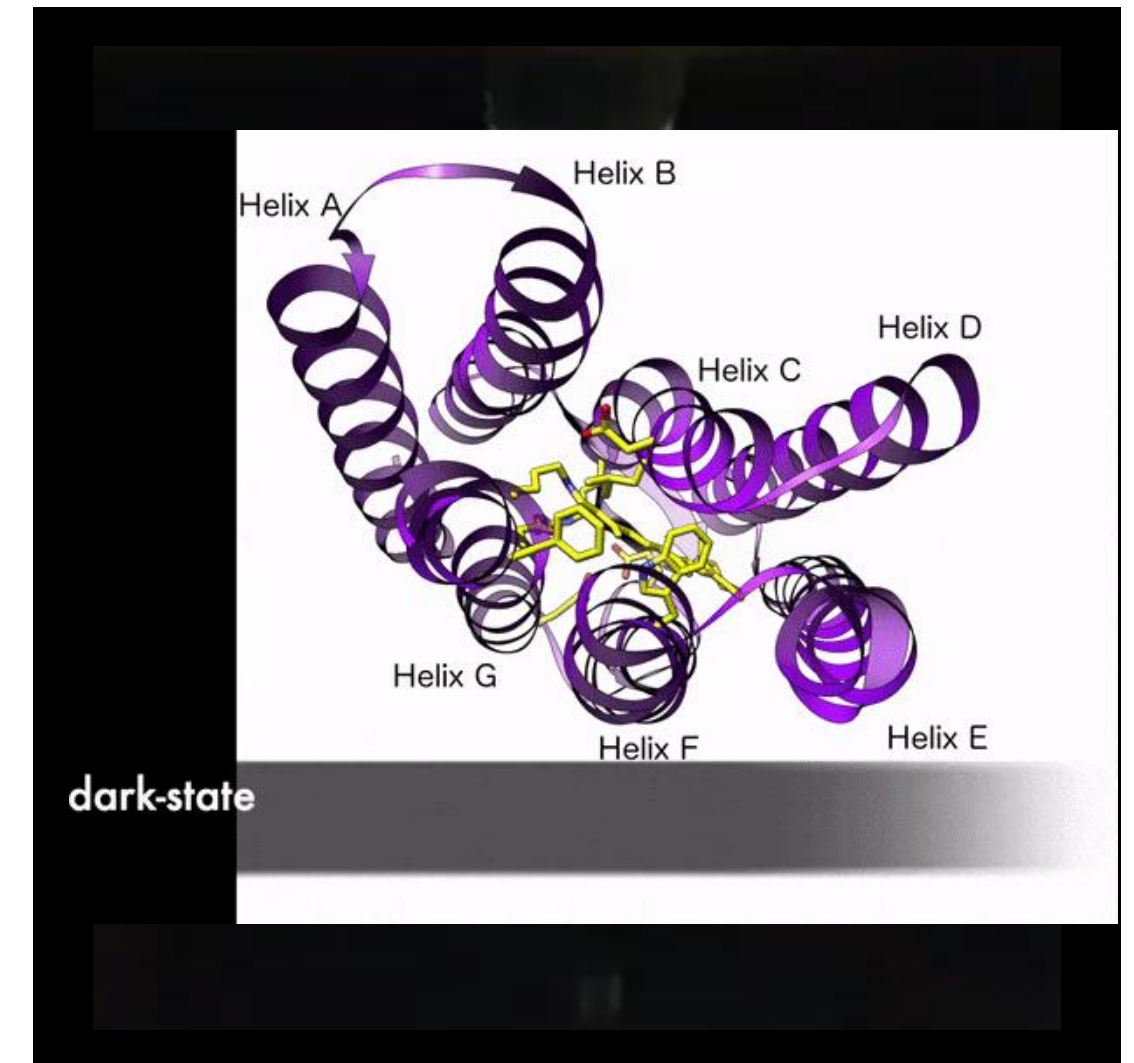
A trigger

Something that starts the reaction on command — a laser pulse for photoswitches, a rapid mix for enzymes.



A fast X-ray pulse, or a fast detector

Pulses as short as femtoseconds freeze each moment like a strobe. One pulse, one snapshot.

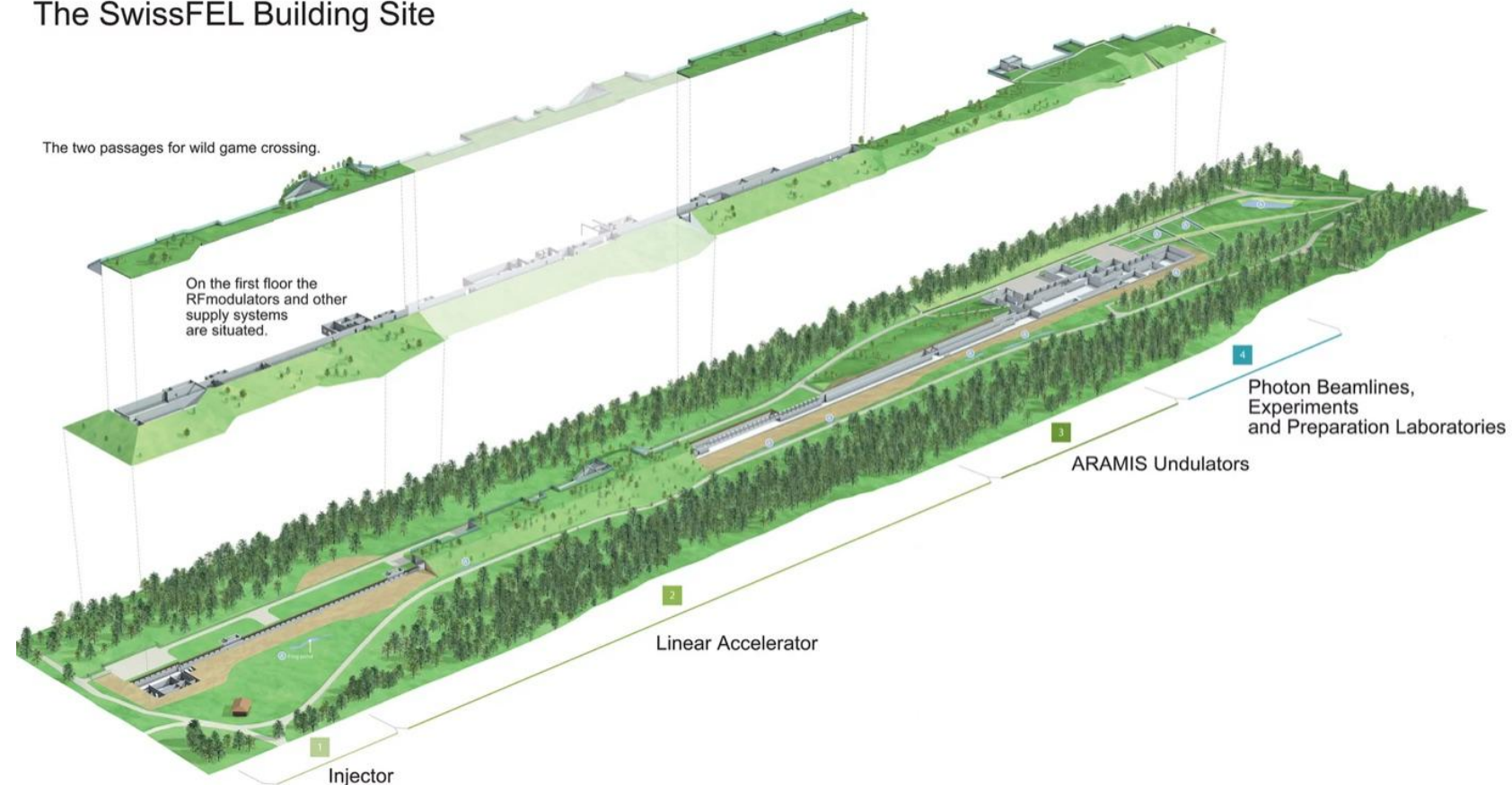


Many crystals

Each crystal gives one shot — destroyed by the pulse at an XFEL, displaced out of the beam in serial millisecond crystallography. Thousands of fresh microcrystals flow through.

Two X-ray sources, two regimes

The SwissFEL Building Site



X-FEL • SwissFEL / SACLA / EuXFEL

Time resolution	femtoseconds
Shots per second	30–120 Hz, or MHz — scarce beam time
Best for	Ultrafast photochemistry, irreversible reactions

4th-gen synchrotron • SLS 2.0 / ESRF / ALBA (soon) / MAX IV

Time resolution	milliseconds
Shots per second	High throughput — abundant beam time
Best for	Slower processes, routine serial crystallography

Not many proteins can be triggered by light. How do we “fix” that?

THE “EASY” WAY

A photoswitch · tubulin · femtoseconds to milliseconds

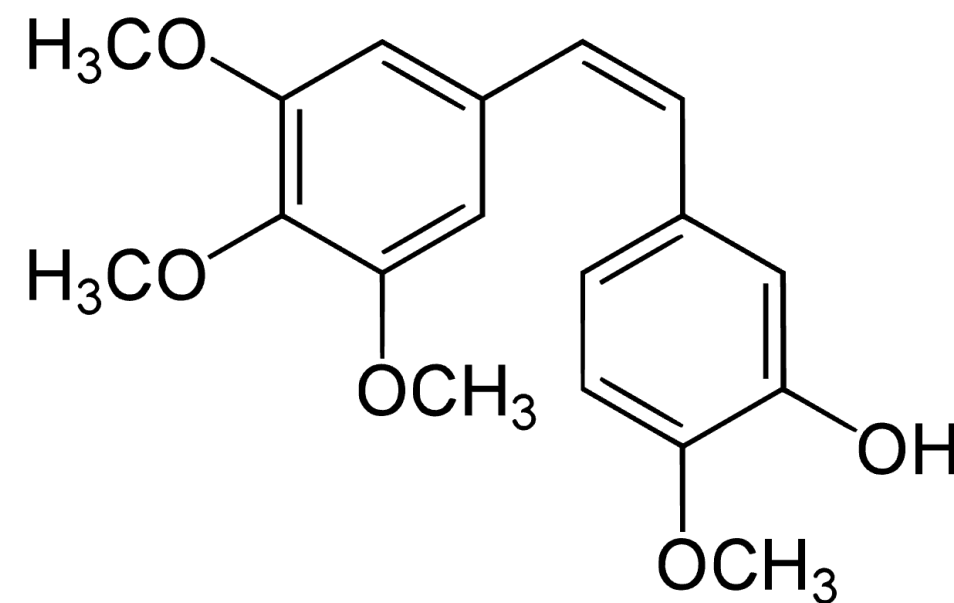
From bark to photoswitch — a cancer drug switched on by light

Replace the C=C bridge of combretastatin A4 with an azo group and you get **azo-CA4**: OFF in the dark (*trans*, inactive), ON at $\lambda = 350\text{--}430\text{ nm}$ (*cis*, binds the colchicine site), and back at $\lambda = 450\text{--}530\text{ nm}$ or spontaneously.



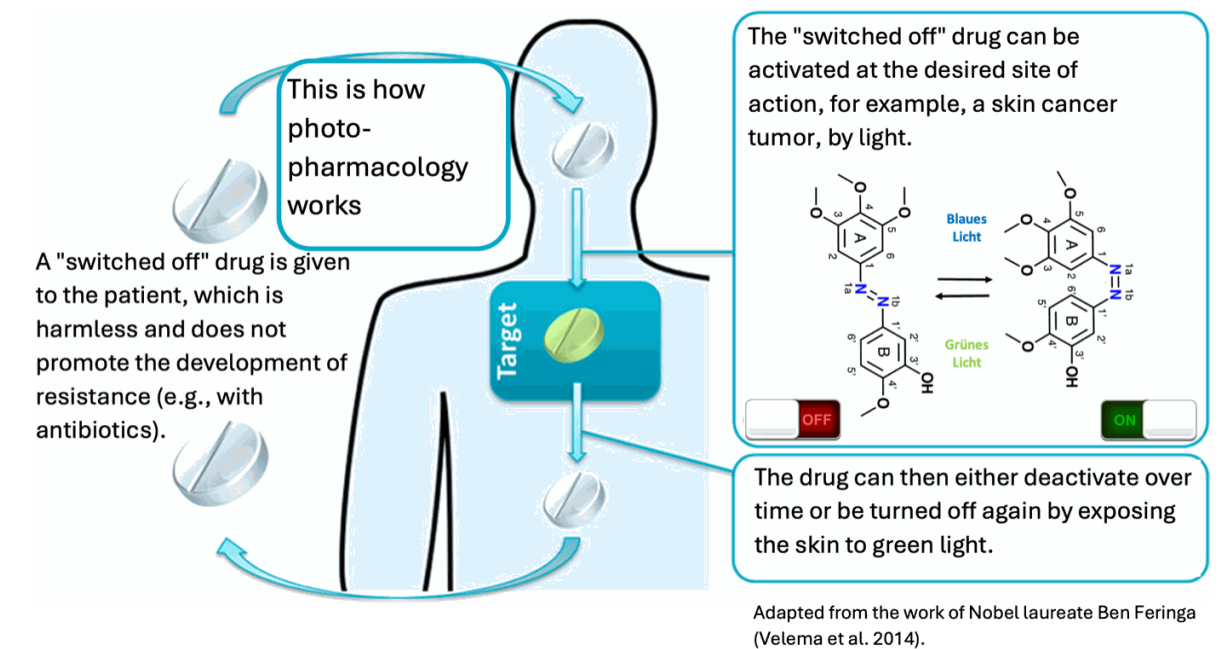
From the bark

The African bushwillow, *Combretum caffrum* — the natural source of the parent compound.



Combretastatin A4

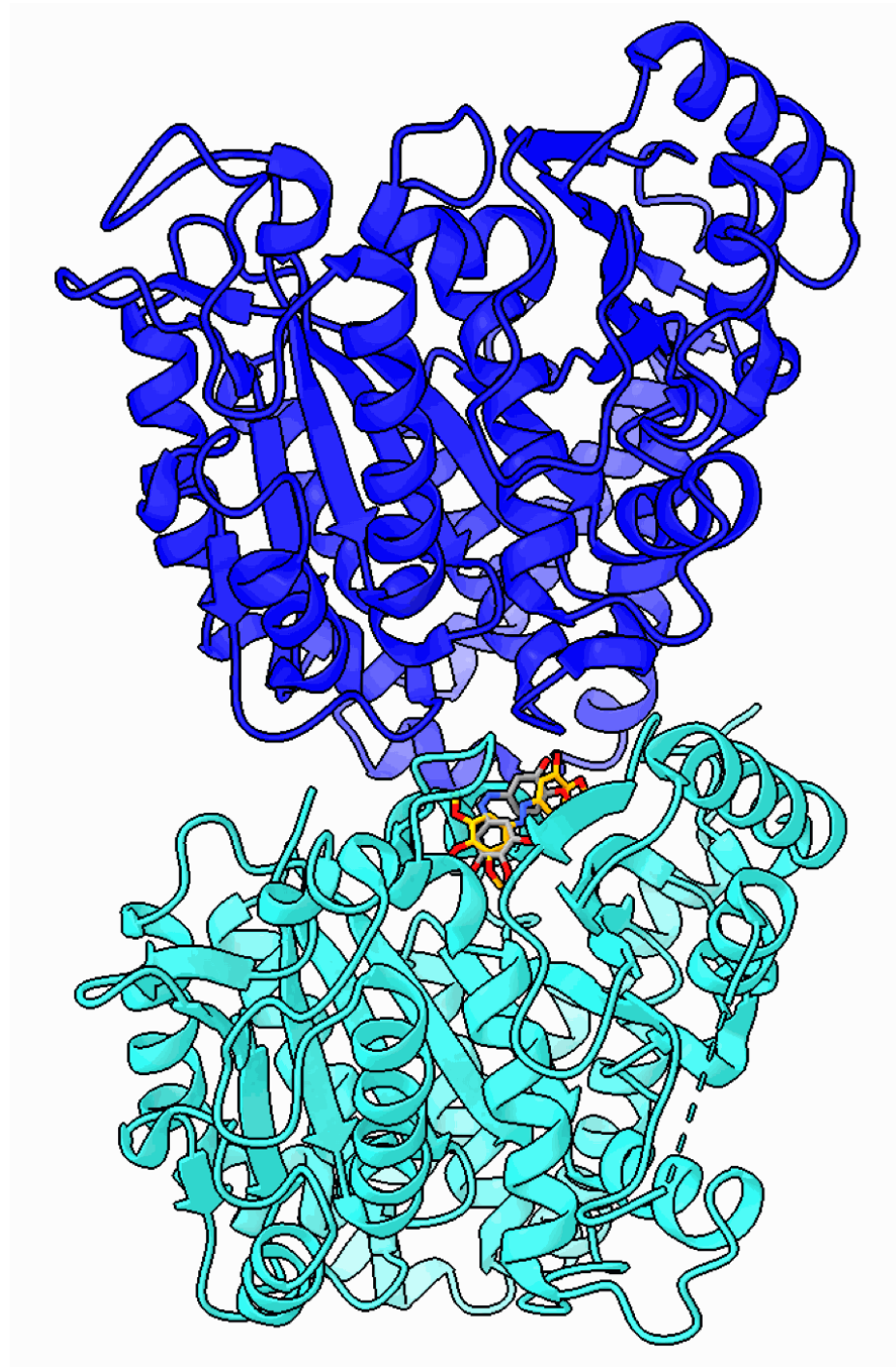
The parent natural product: a potent tubulin binder at the colchicine site, and the scaffold the azo group replaces.



Why it matters

The drug is only active where the light is. Toxicity stays out of the rest of the patient.

From photopharmacology to time-resolved serial crystallography



Green light resets every crystal to *trans* before it reaches the beam. The pump laser fires, the delay Δt is scanned, and each time point is merged from thousands of crystals.

DELAYS MEASURED

–500 fs · –230 fs

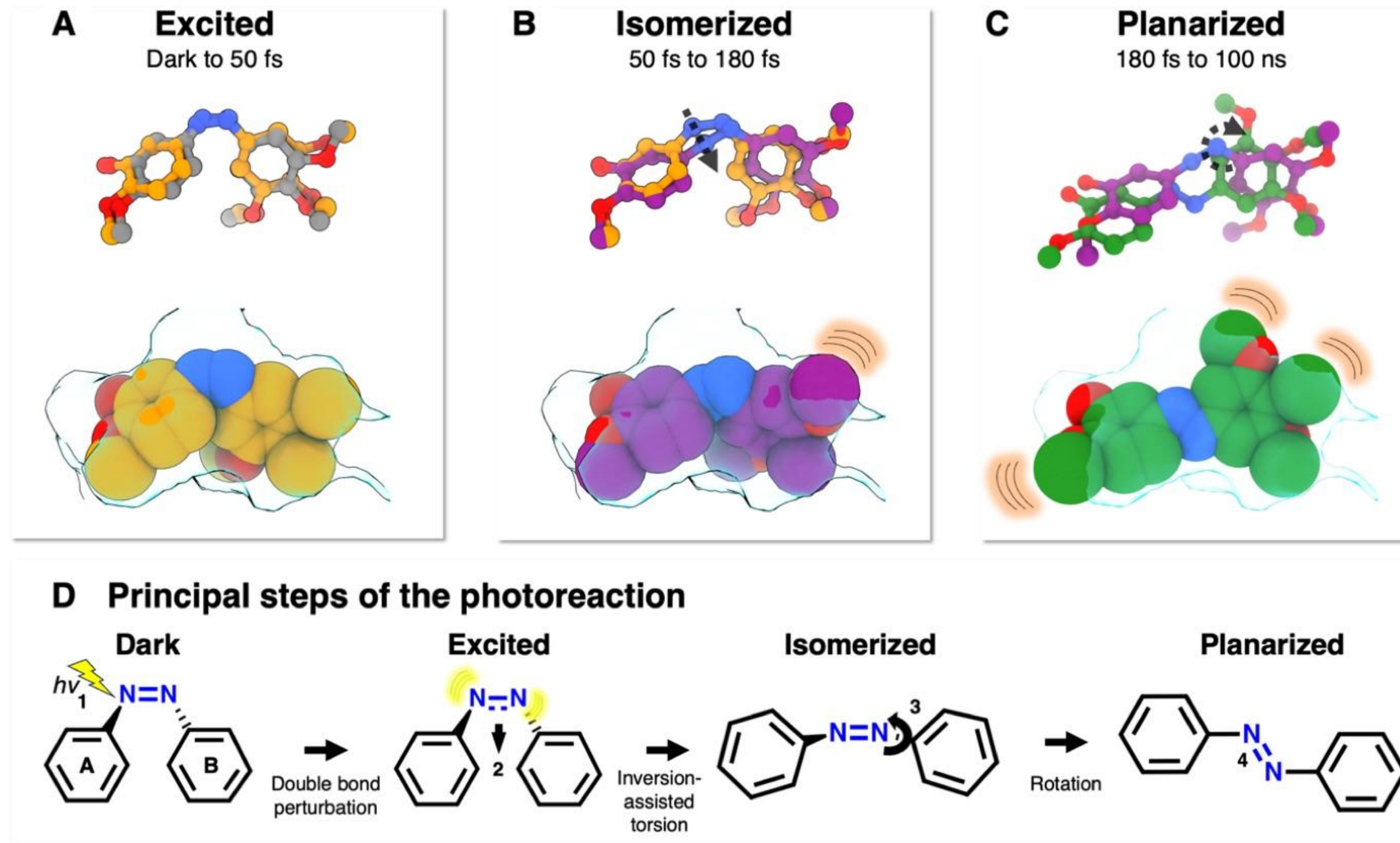
450 fs — the fs scan

1 ps · 25 ps · 35 ps

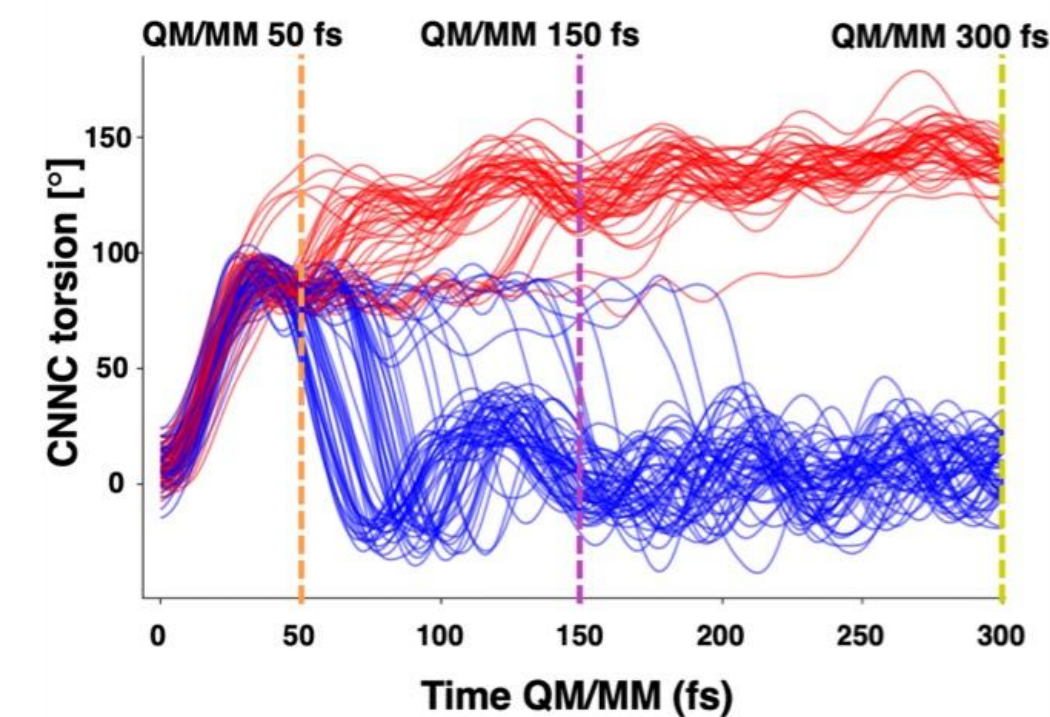
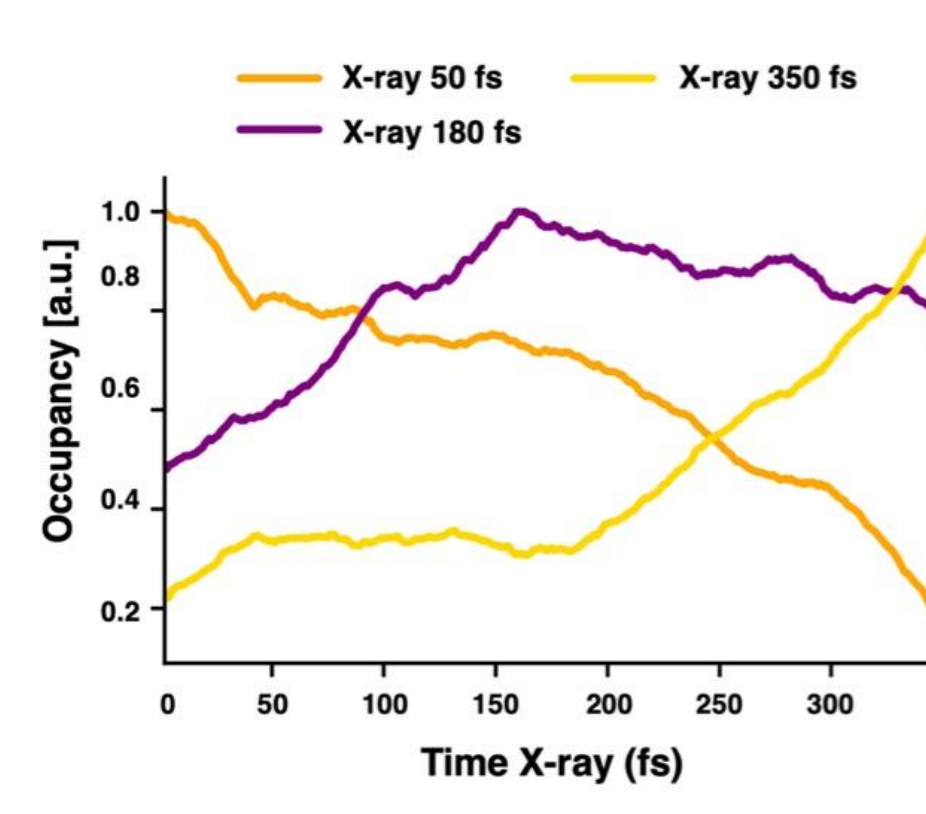
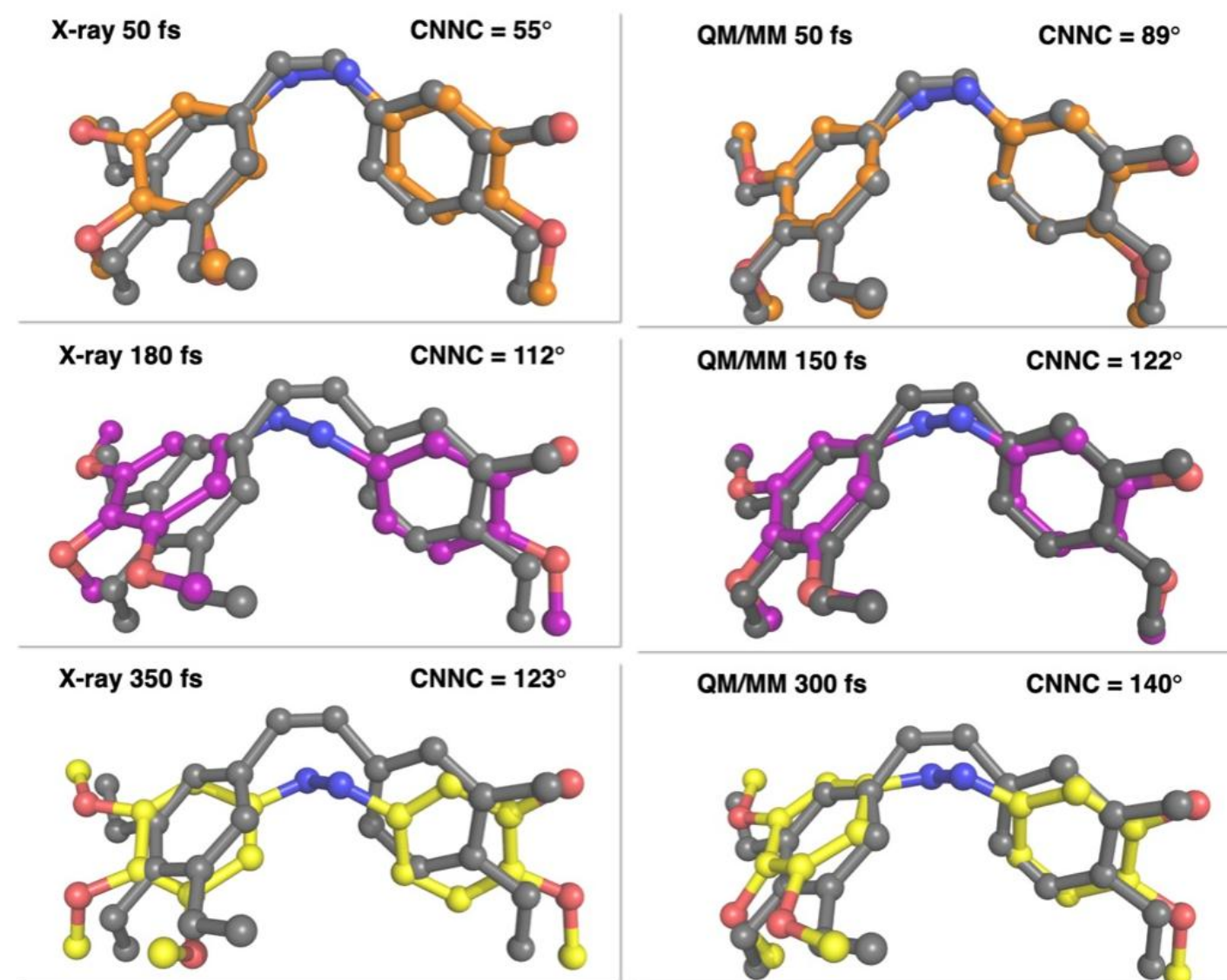
125 ps · 1 ns · 10 ns

100 ns

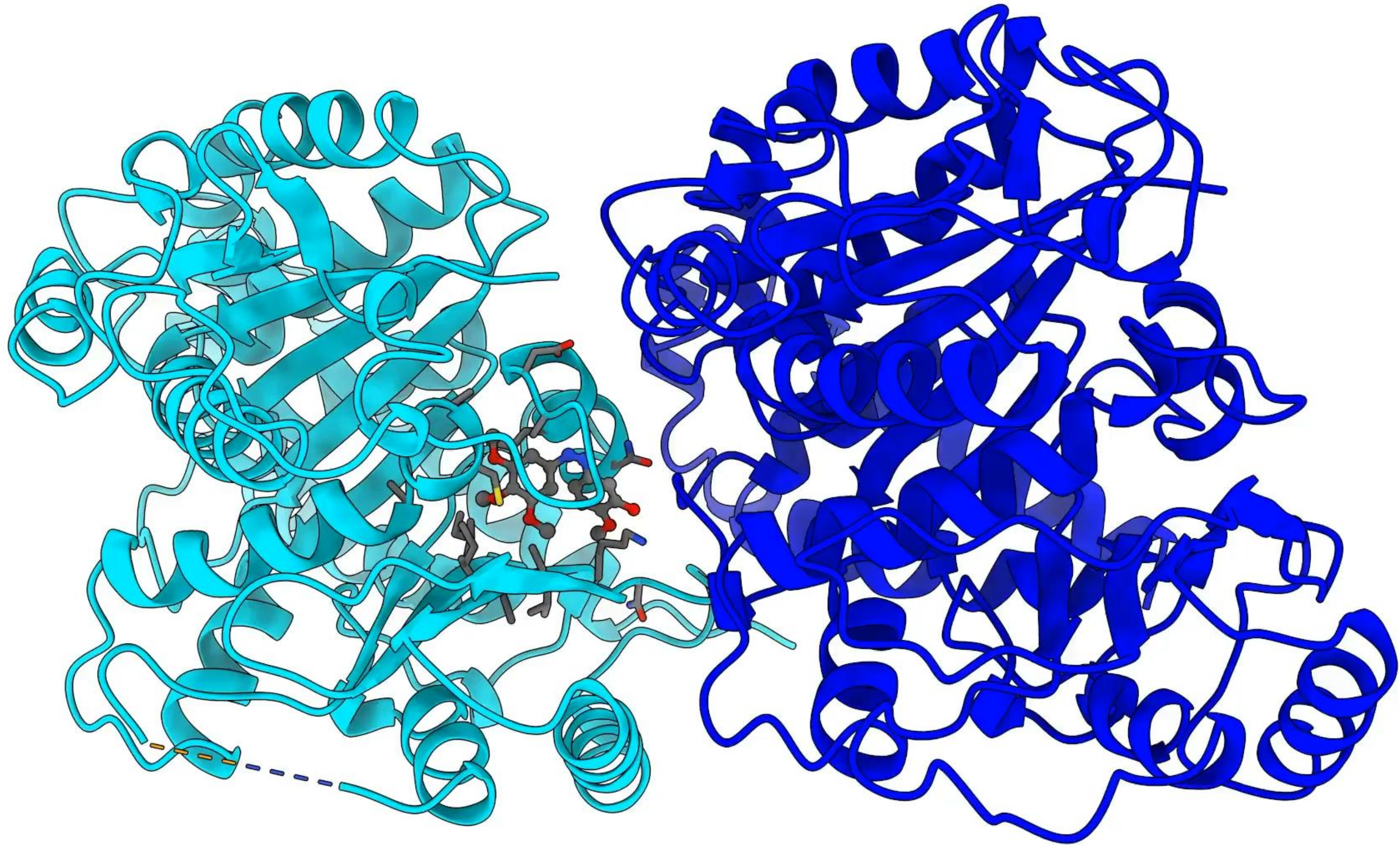
Chemistry — azobenzene isomerising inside a protein, femtoseconds to nanoseconds



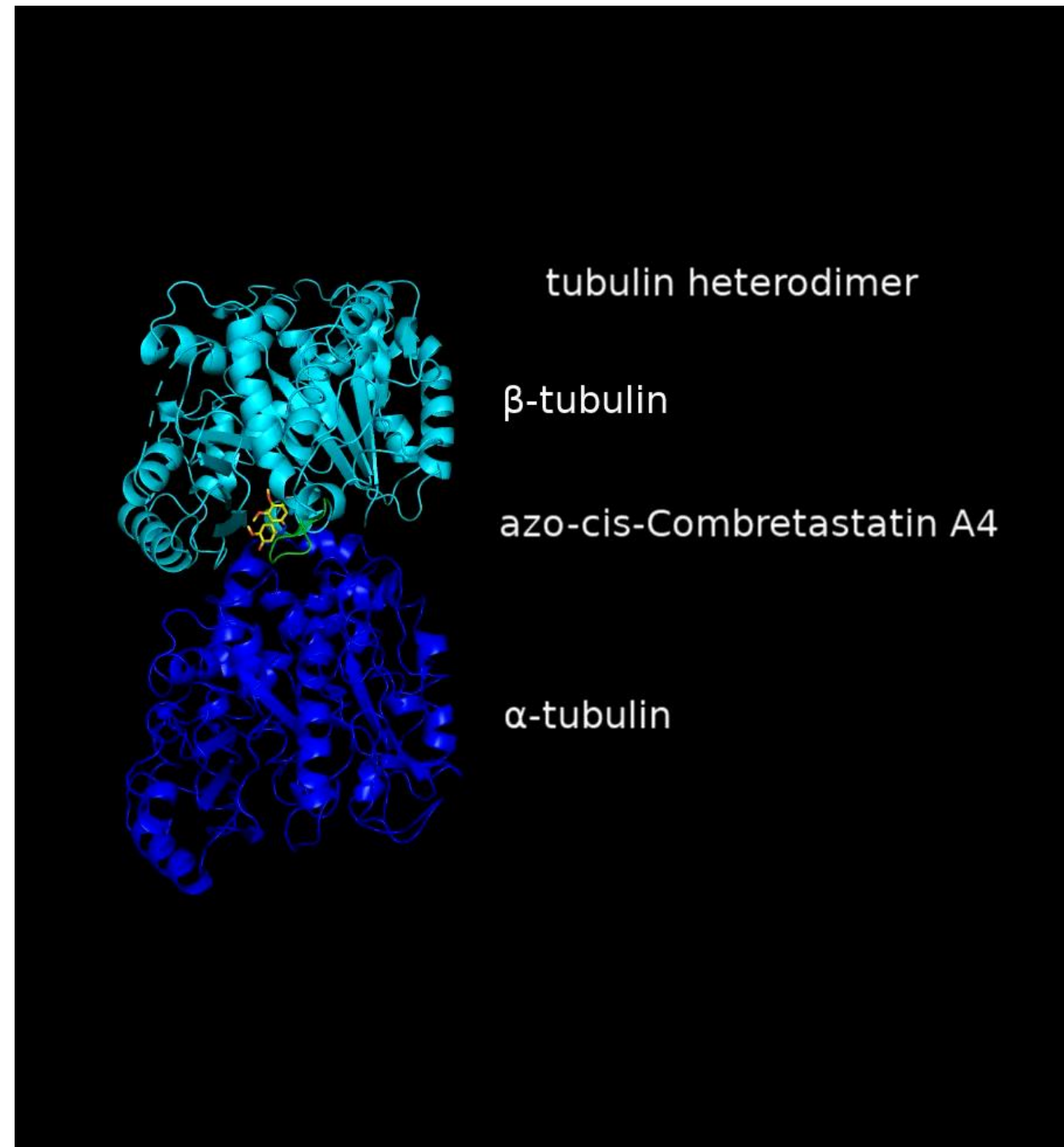
The femtosecond scan — states are already convoluted



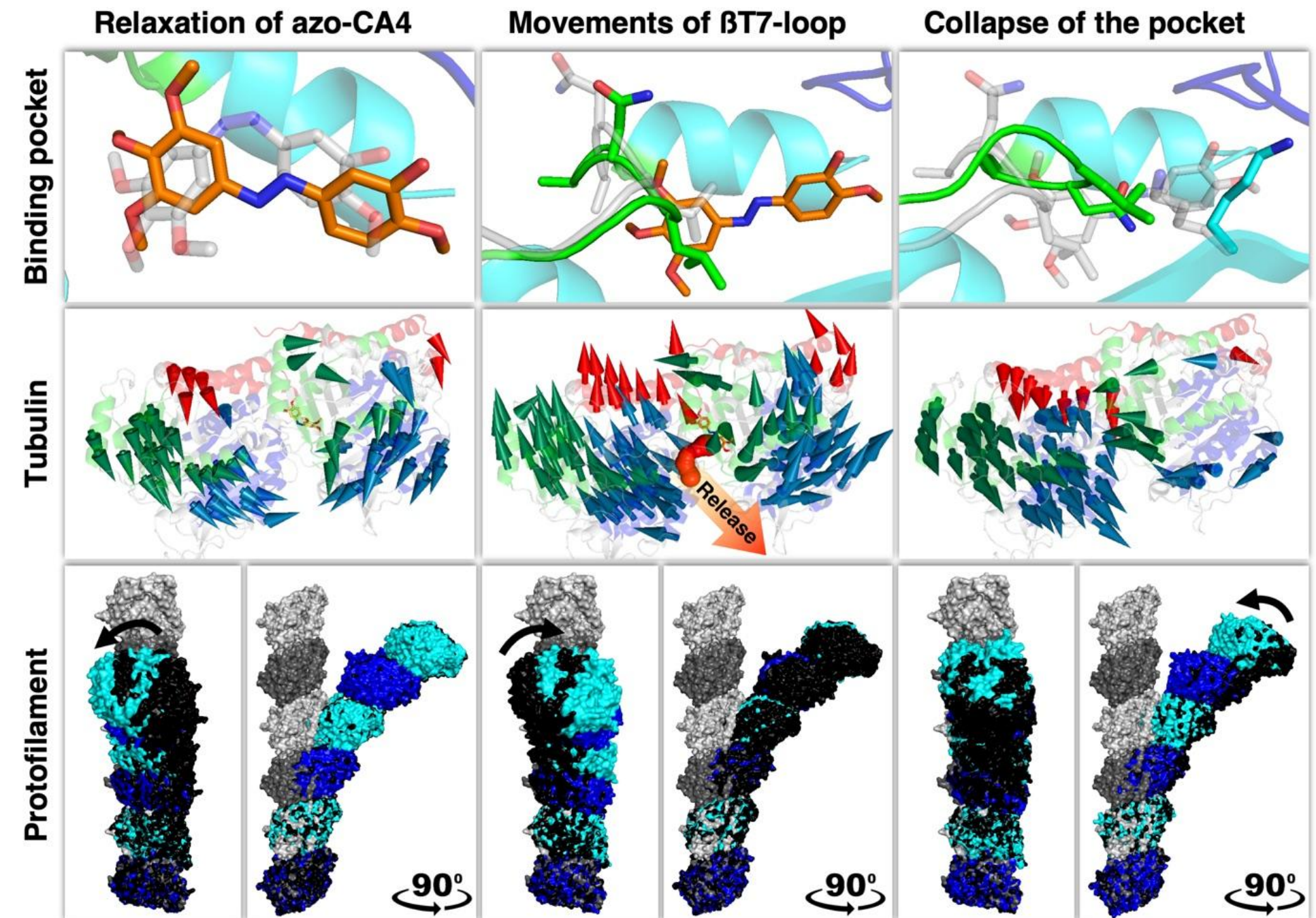
Watching Azobenzene Isomerization Inside a Protein



Pharmacology — watching a ligand unbind from its target



Wranik et al. 2023



What we gain

- Femtosecond snapshots
- Atomic resolution at room temperature
- Undamaged structures
- Full reaction trajectories

What it costs

- XFEL beam time is extremely scarce
- Millions of crystals per dataset
- Complex sample delivery
- Data volumes of terabytes per hour

And it still assumes the reaction has a trigger, and that one state is present at a time. The next system has neither.

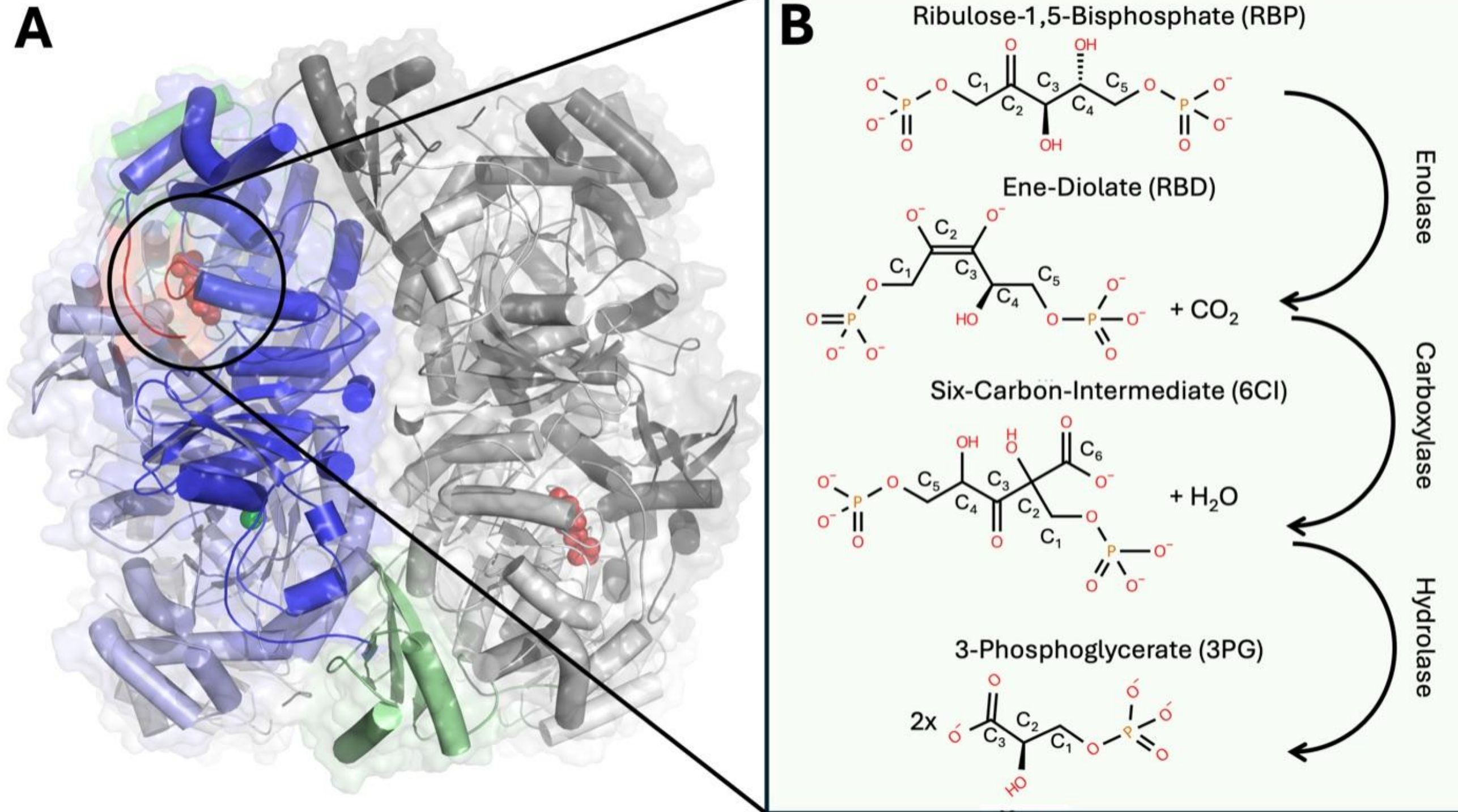
RuBisCO

THE “HARD” WAY

Four reactions · convolution of states · no trigger

Most of this work was done by **Georgii Khusainov**, who is in the room today.

RuBisCO — the most important enzyme on Earth

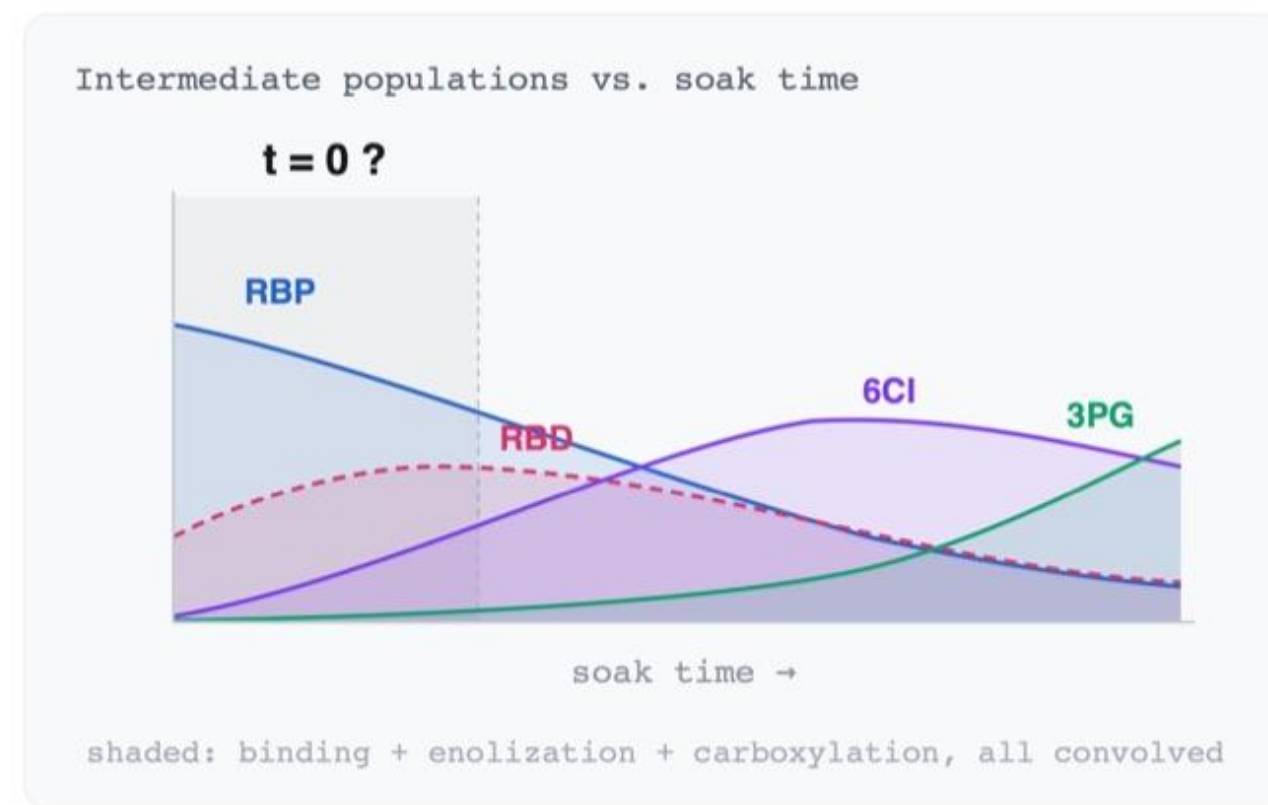


Convolution of states — usually ignored in crystallography

Four reactions run in series, and none of them waits for the last to finish. At no point in the time course is a single intermediate alone in the crystal — so every map is a weighted sum of states, and refining one model into it gives an average that matches nothing.

■ THE LIMITATION

The bottleneck: intermediates overlap, with no clean $t = 0$



01

States populate as mixtures

Every dataset is a superposition of intermediates — structures only emerge after heavy deconvolution.

02

No defined start, no tunable rate

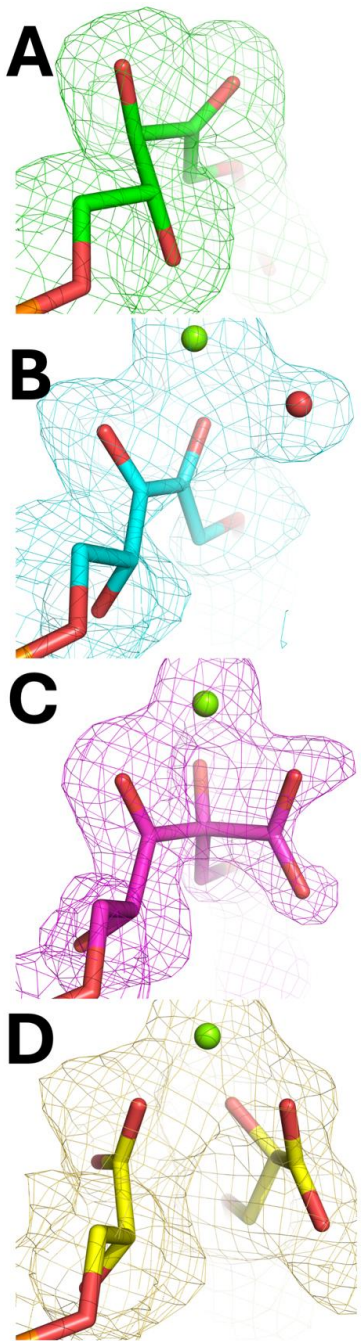
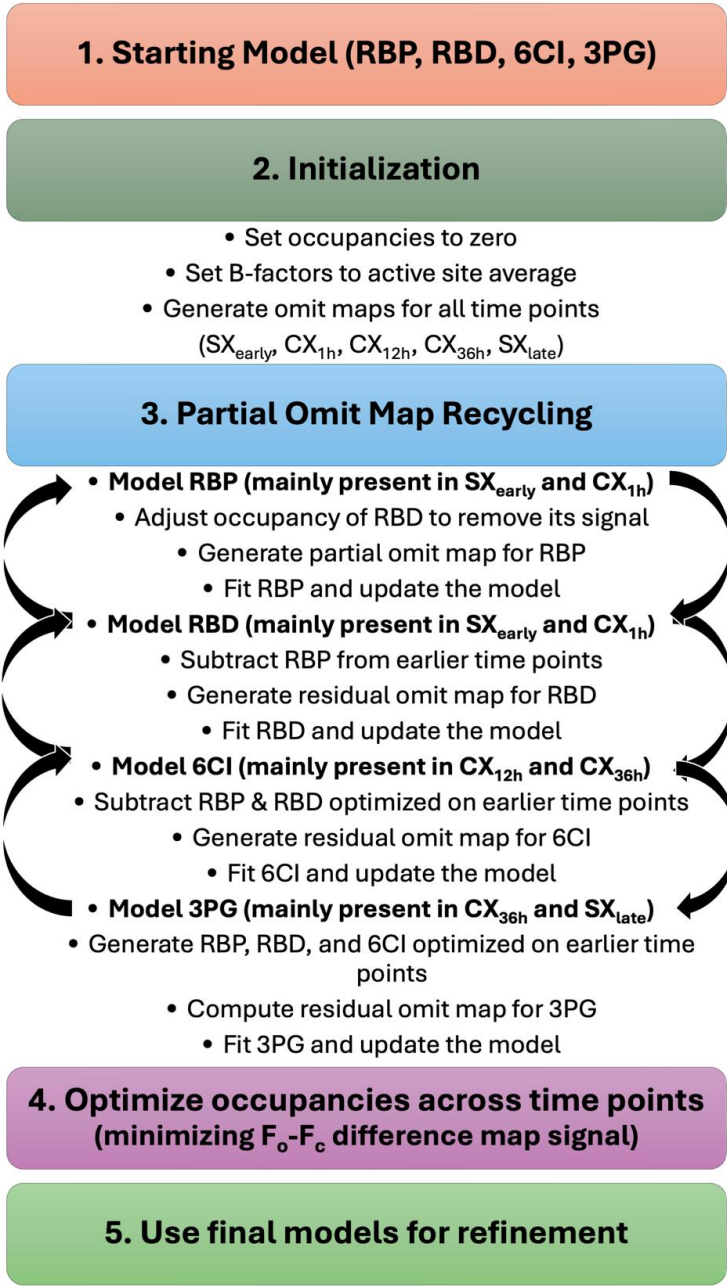
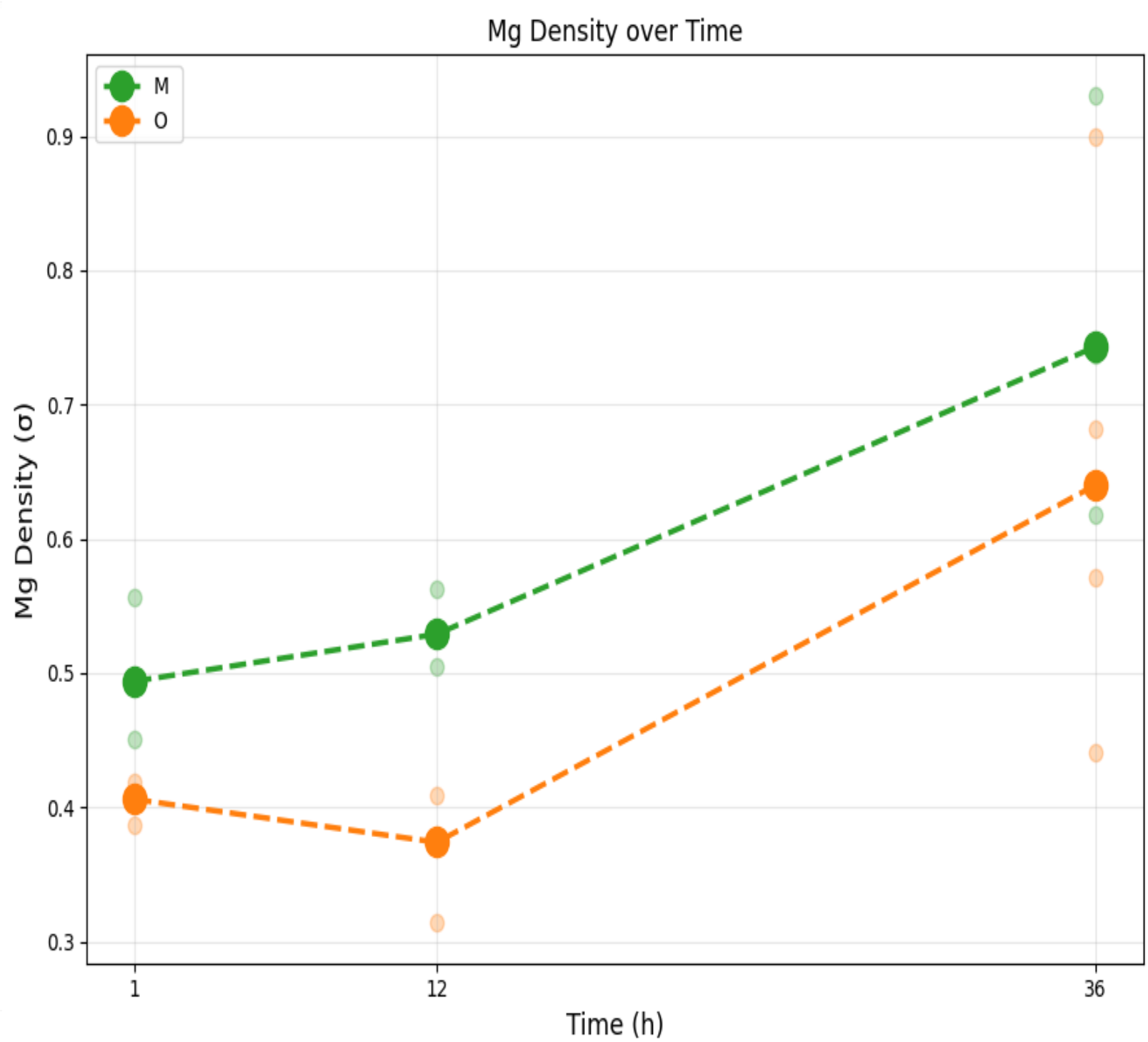
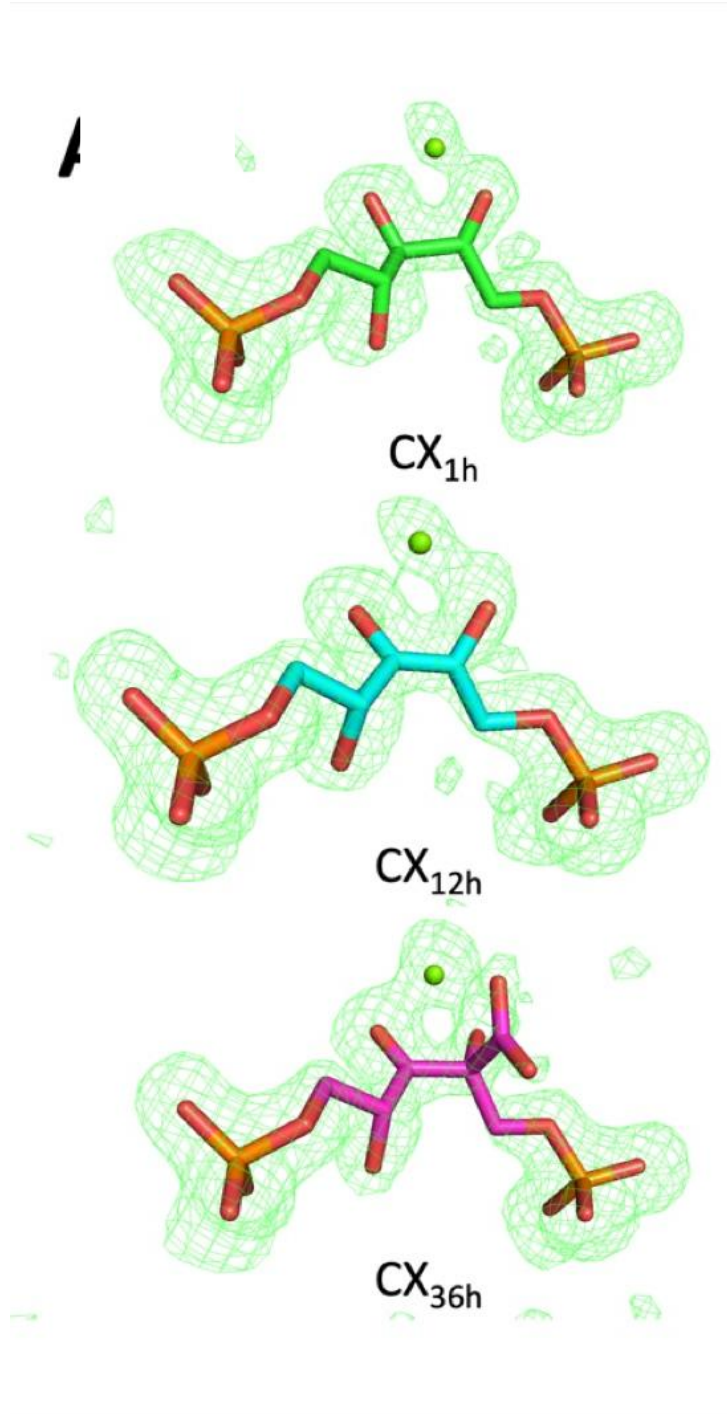
Binding, Mg^{2+} expulsion and carboxylation onset all overlap; $t = 0$ is indeterminate.

03

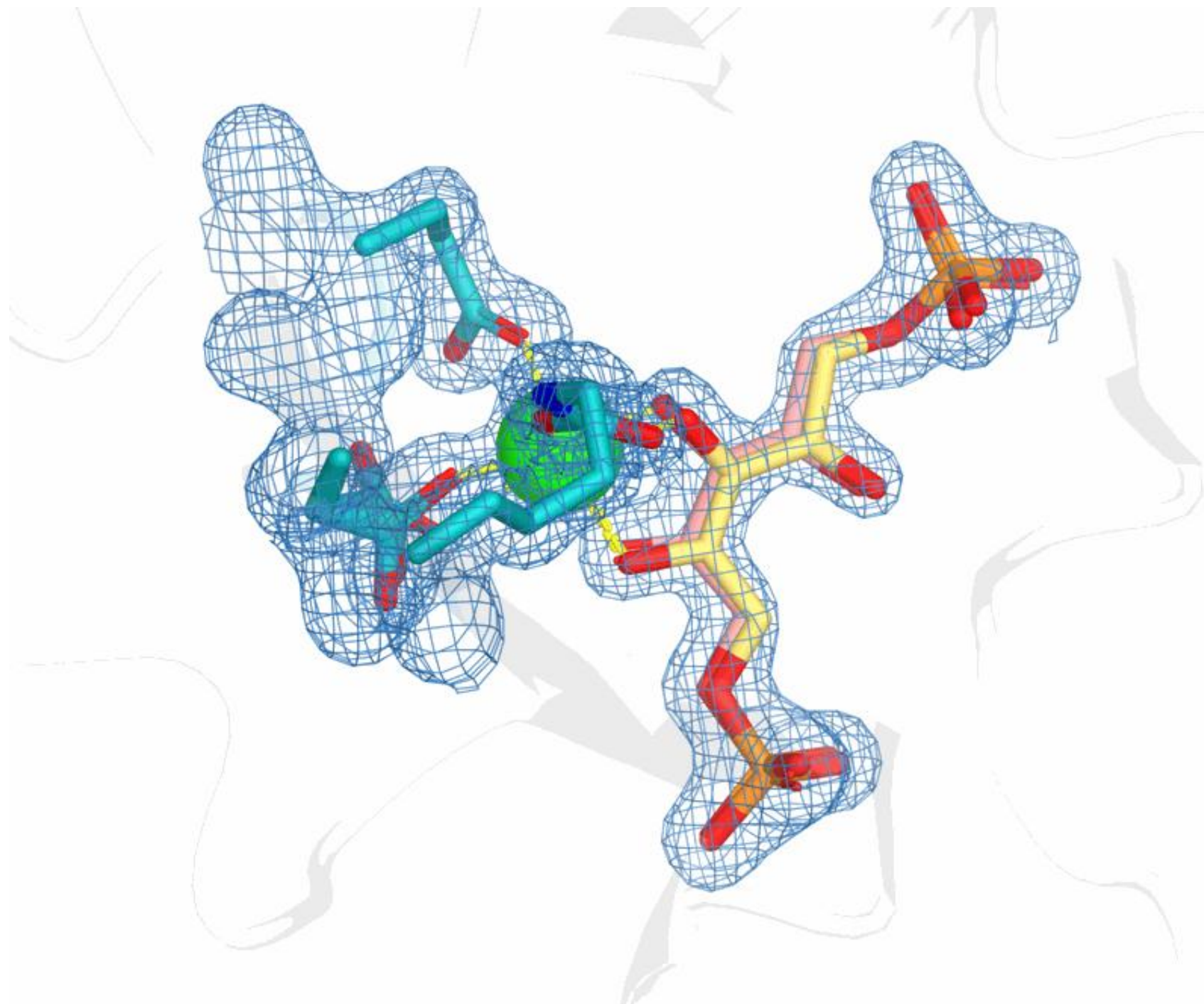
The ene-diolate stays hidden

RBD never exceeds ~40% occupancy — the pivotal catalytic state is the least certain of all.

Mg²⁺ leaves — and comes back



A trapped ene-diolate — and a coordination that did not fit



WHAT THE T-JUMP GAVE US

The difference maps were **flat**. The temperature jump did not drive the reaction.

WHAT THE DENSITY GAVE US INSTEAD

Two sp^2 -hybridised carbons — the ene-diolate, the species we had already untangled from the cryo series. We were pleased with ourselves.

THEN THE METAL

Here

5-fold

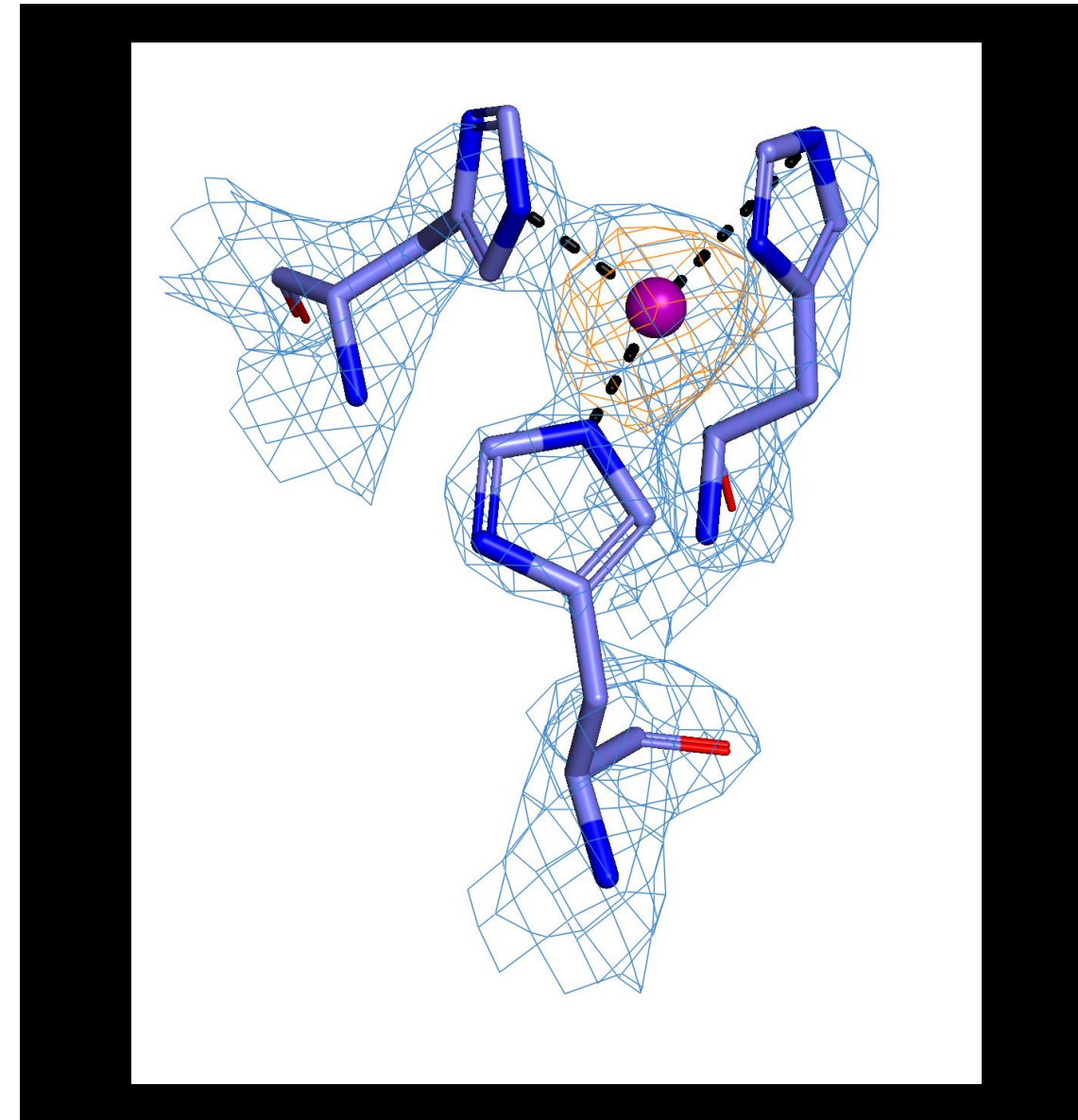
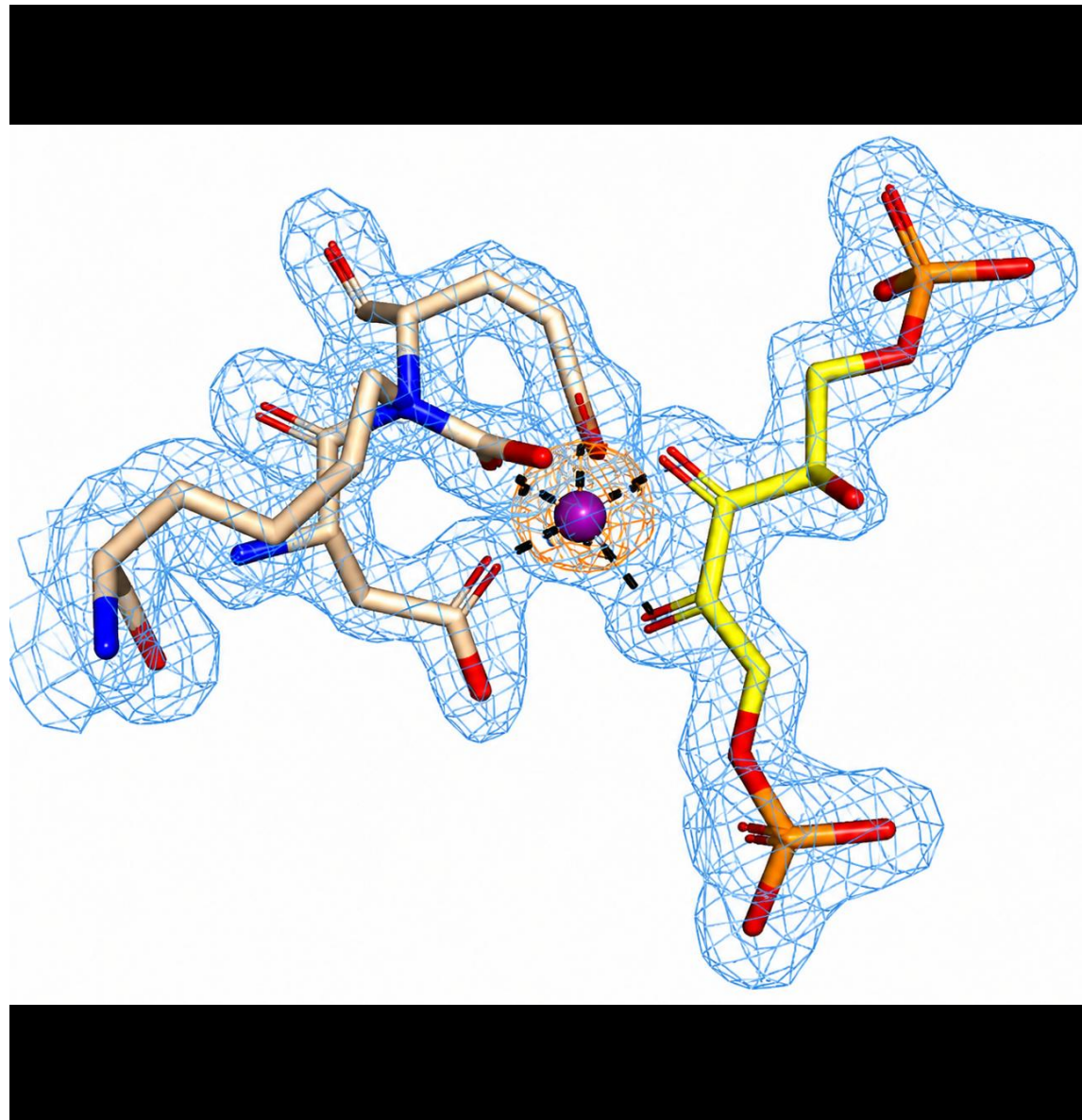
The coordination did not match. So we went looking for alternative theories.

Cryo model

6-fold

The alternative theory: Zn^{2+} — contaminant, or contributor?

We were above the zinc edge — and found a very strong anomalous peak **at the metal site**, which accounts for the five-fold coordination.



Two sites, not one. The strong peak sits at the metal site. In the subunits *not* occupied by substrate, a second peak appears at a nearby histidine cluster.

ESRF, without substrate, found that same adjacent site — there the fourth ligand is the Lys-carbamylate. We are still making sense of it.

The technique is not yet routine.

ANALYSIS

PROBIXI

ACQUISITION

HATRX

CAN IT BE DONE AT ALL?

DeconX

REFINEMENT

TorchRef

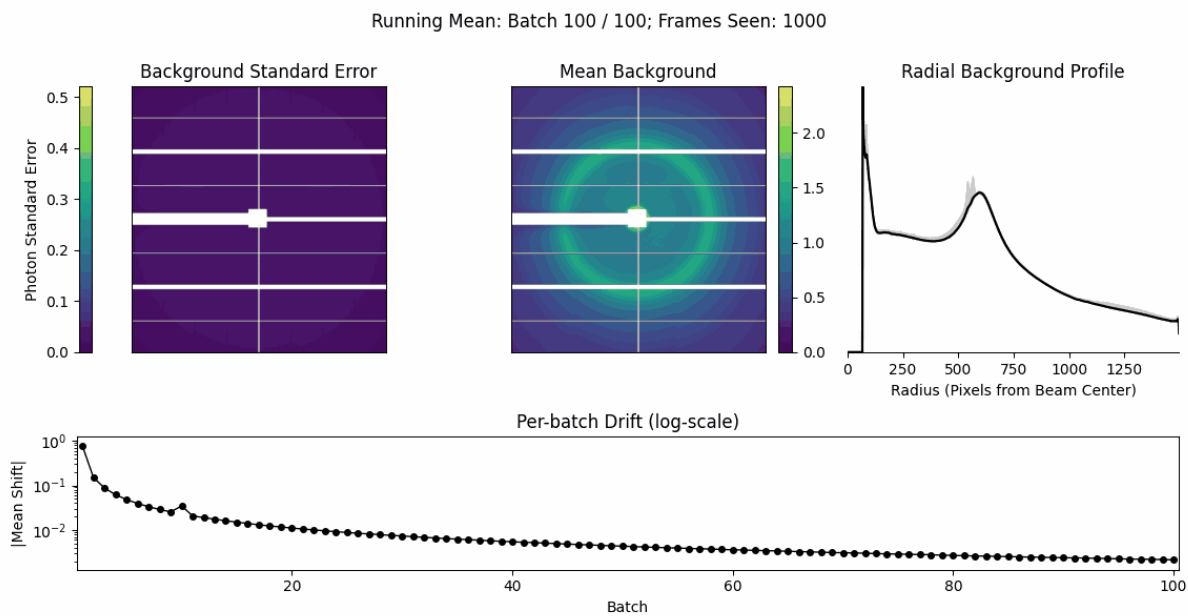
PROBIXI — peakfinding as anomaly detection against a learned noise prior

Learn per pixel what quiet looks like over a few seed frames, whiten the frame into a z-map, then force that map to be normal by searching the blend of pixel, radial and panel statistics that makes it so. Peaks are what is left over.

ONE PIECE OF MATHS

$$\log \text{BF} = -\log \kappa + \frac{1}{2} \mathbf{z}^2 \left(1 - \frac{1}{\kappa^2}\right)$$

Peak contrast κ , the prior, the mixture weights and the detection threshold are all solved on the seed sequence — none of them is tuned by hand.



C1C2 CHANNELRHODOPSIN · 8.72 MILLION FRAMES · 2.60 Å

METRIC	pf8 + XGANDALF	PROBIXI subset	PROBIXI
Indexed frames	3.8×10^4	3.8×10^4	2.4×10^5
Indexing rate (%)	0.44	—	2.30
R_{split} (%)	21.83	11.73	5.08
$\text{CC}_{\frac{1}{2}}$	0.979	0.980	0.999
$\langle I/\sigma(I) \rangle$	2.04	4.90	13.54

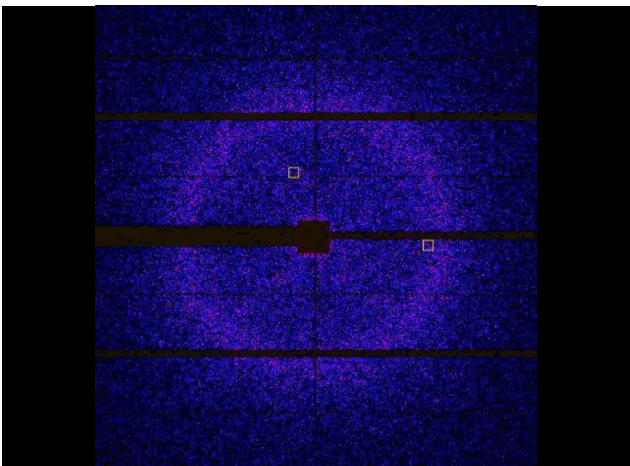
Merged with `partialator` under the `xsphere` partiality model; 2.60 Å is the nominal $\text{CC}_{\frac{1}{2}} > 0.3$ limit reached by both pipelines. The **subset** arm caps PROBIXI at the same frame count as the tuned pipeline — the gain is not just multiplicity.

Highest shell, R_{split}	Wall clock per crystal	Parameter tuning
58× lower	0.87 s vs 8.82 s	0 h vs 89 h

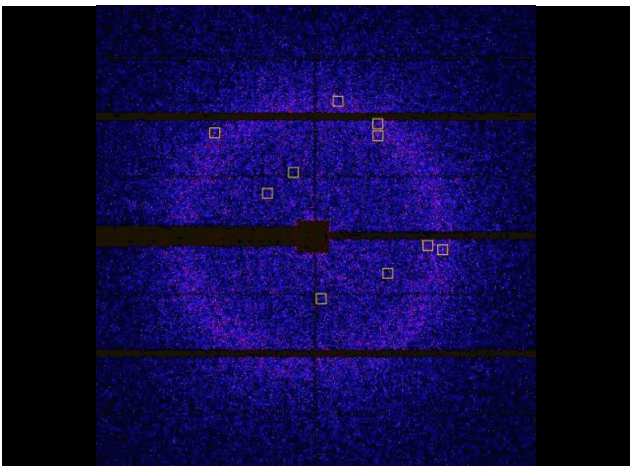
HATRX — encode time points before indexing, decode after merging

$$\begin{matrix} w_1 & 1 & 1 & 0 & I_1 \\ w_2 & = & 1 & 0 & 1 & \cdot & I_2 \\ w_3 & & 0 & 1 & 1 & & I_3 \end{matrix}$$

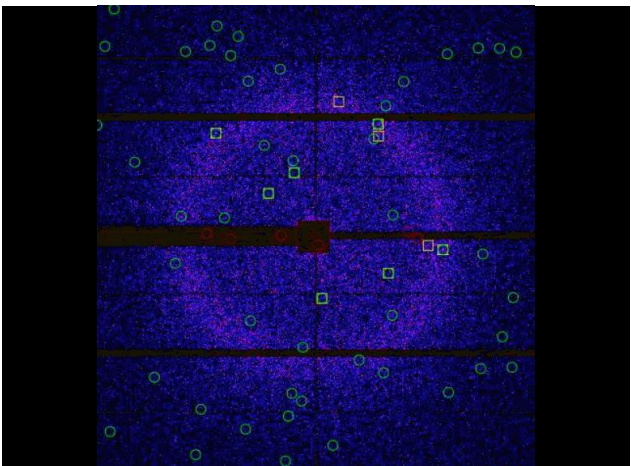
Sum by the pattern **S** before indexing;
decode the merged intensities with **S**⁻¹.
Detector gating gives time resolution
faster than the frame rate.



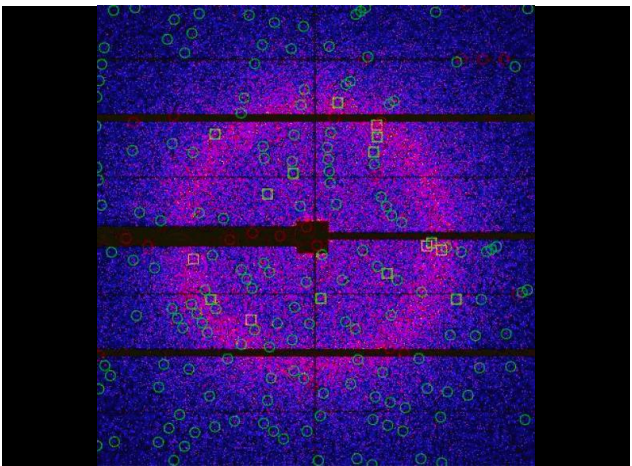
Will not index alone



Will index alone



Indexed conventionally



The HATRX sum indexes — both
time points now contribute

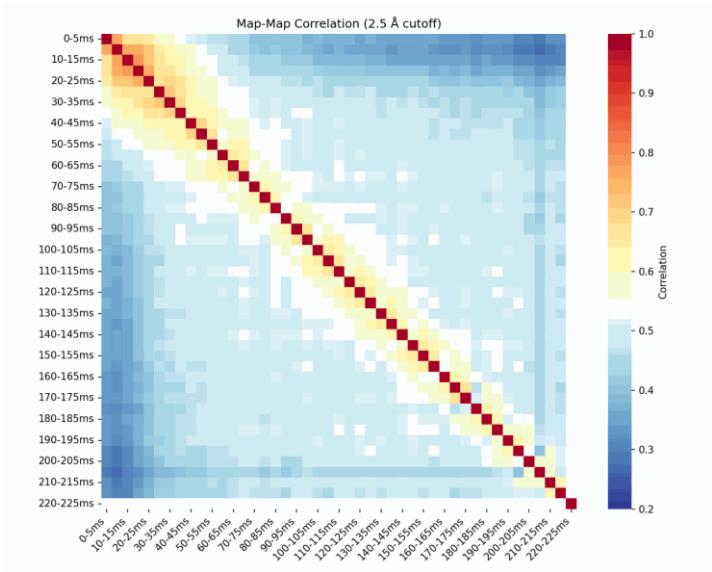


Fig. 18 — bR map–map correlation under HATRX, no scaling.

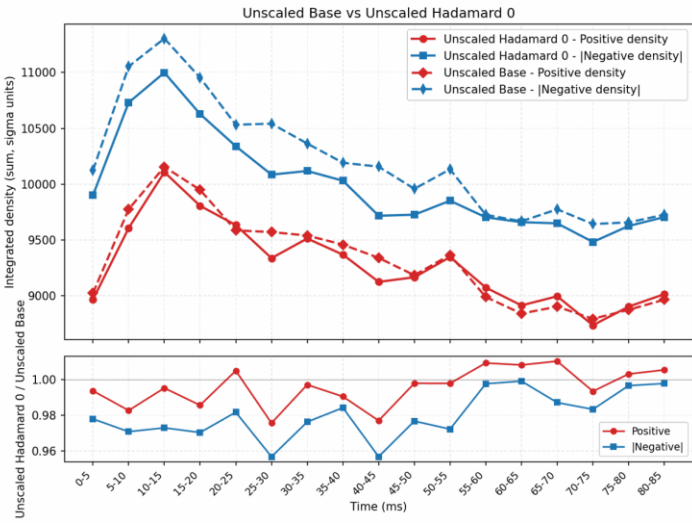


Fig. 19 — Signal along the opening helix, bR, unscaled: HATRX against canonical.

98 %

of the bR signal recovered where signal is already rich — and it begins to recover *more* where signal is poor.

A proof of principle: HATRX **recovers** the signal, it does not improve it. The gain is at low signal-to-noise — it is not a free lunch.

Can You Deconvolve States From Difference Electron Density Maps?

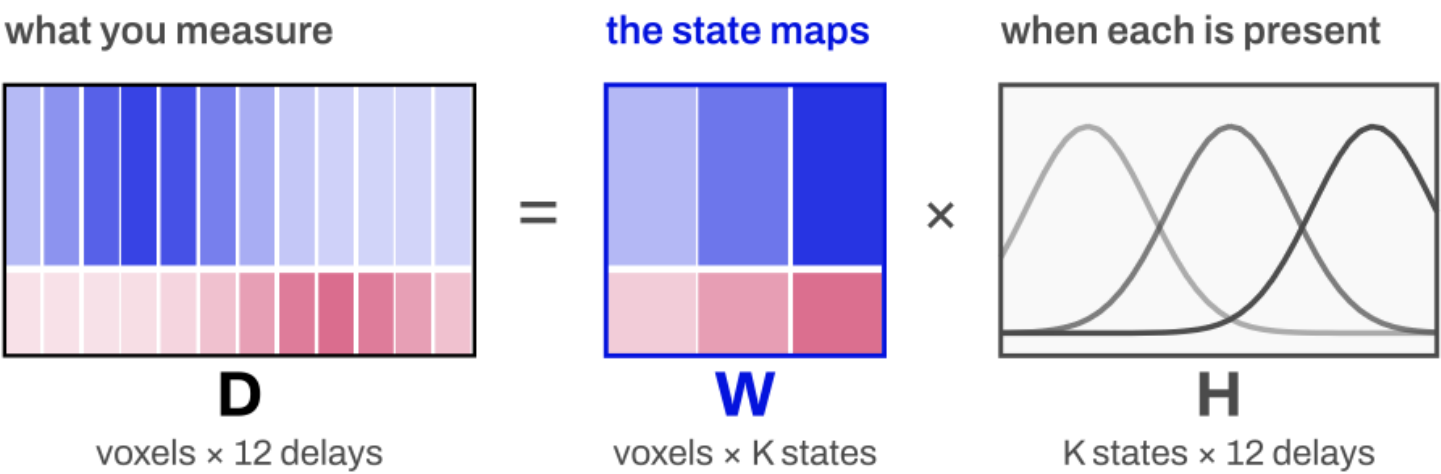
$$D = W H + E \quad H \geq 0, \quad \sum_k H[k, t] \leq 1$$

Build the mixtures from deposited PDB models so that **W** and **H** are known exactly. Then every unmixing method gets a number instead of an argument.

THE CORPUS — PDB AS OF 27.08.2026, 66 GATES OVER 5 TIERS

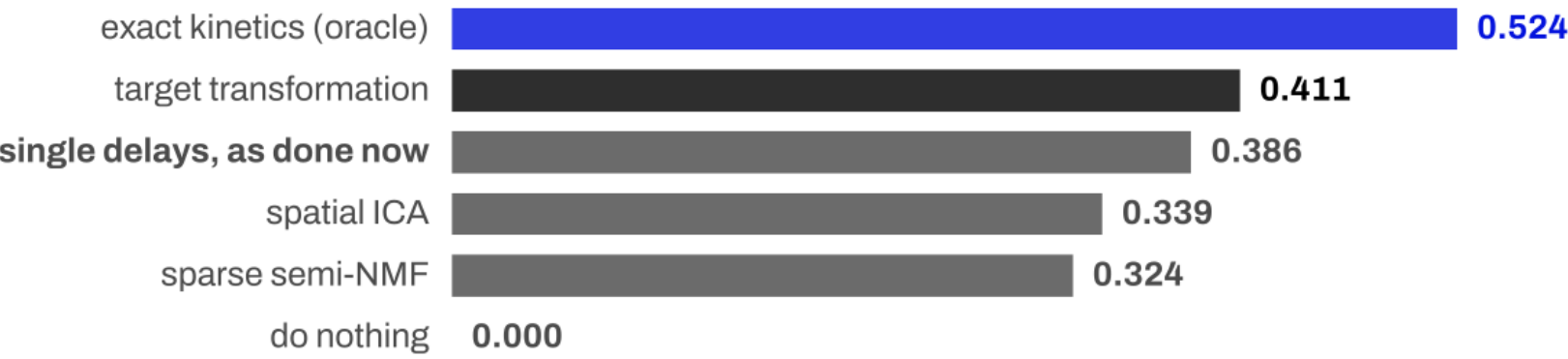
X-ray entries examined		162 846
Pass entry quality		90 583
Crystal-form families		6 791
Families that can pose 2+ states		2 301
Experiments passing every gate		37 425

185 distinct proteins · 426 crystal forms · $\approx 2.0 \times 10^6$ experiments supported. Bar length is log-scaled.



Deconvolution: recover **W** and **H** from **D** alone. Both are known here.

No blind method wins



TorchRef

An open-source PyTorch framework for crystallographic refinement



H. P. Seidel · J. Standfuss · T. Weinert

<https://doi.org/10.64898/2026.05.13.724821> (Just accepted yesterday)

As accurate as Phenix — orders of magnitude faster



< 1 %

median R-free gap to Phenix across the benchmark

0.3×

median wall-clock of an identical Phenix refinement

> 100×

faster structure factors on a GPU than CCTBX

1 000

PDB structures refined end-to-end, 1.0–3.0 Å

BENCHMARKED LIKE FOR LIKE

Perturbed starting models, matched protocols, then re-scored independently in REFMAC5. Geometry and ADP quality sit on par with Phenix.

Structure-factor calculation already beats CCTBX past two CPU threads, and runs more than twice as fast at four cores before the GPU pulls two orders of magnitude ahead.

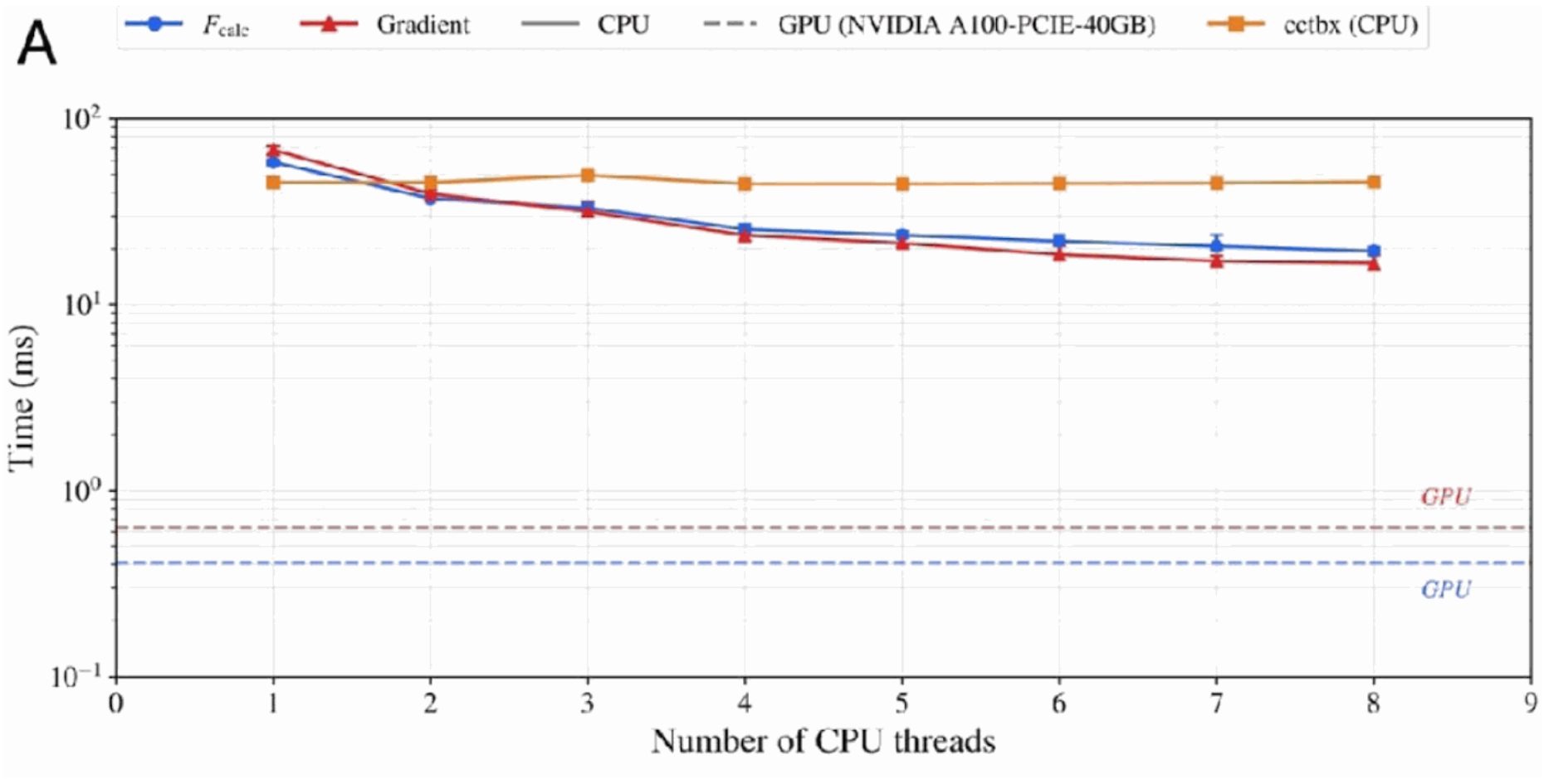


Fig. 20 — Wall-clock per F_{calc} evaluation against CPU threads; dashed lines mark GPU timings (NVIDIA A100).

Refining directly against the difference

End-to-end differentiability lets us fit the change itself — a combined dark + light model refined straight against the observed amplitude differences, with no extrapolation.

THE USUAL ROUTE — EXTRAPOLATED STRUCTURE FACTORS

Amplify a partially occupied state to mimic full occupancy. Noise grows as occupancy drops, dark-state phases leak in, and negative amplitudes need patching — or Bayesian shrinkage that biases back toward the dark state.

TORCHREF — DIRECT DIFFERENCE REFINEMENT

Fit Δ (dark $\rightarrow$ light) against the observed ΔF directly. Systematic errors common to both datasets cancel — and the whole target was about 200 lines of Python.

0.847

vs 0.735 extrapolated

Local correlation to the difference density around the ligand.

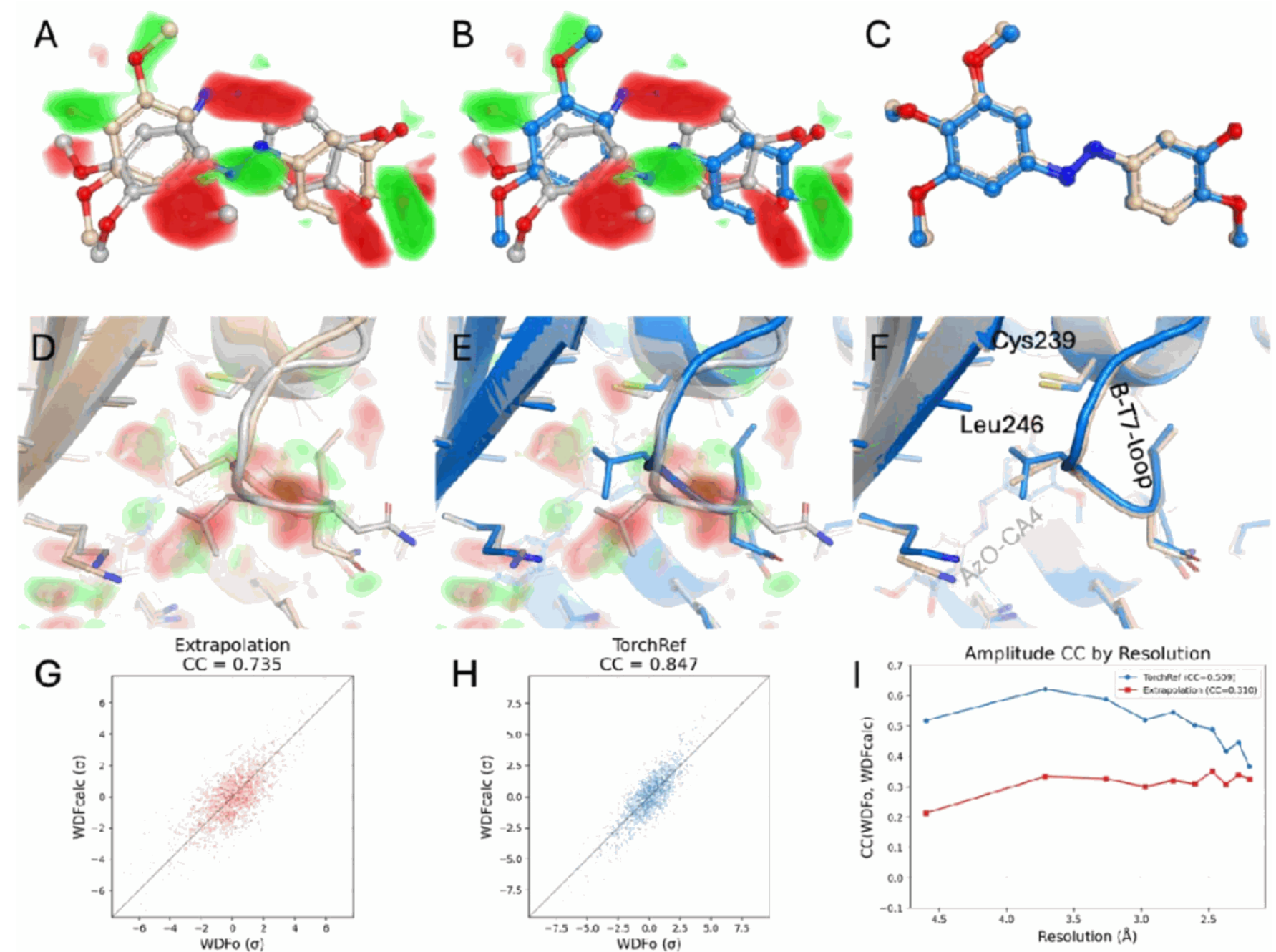
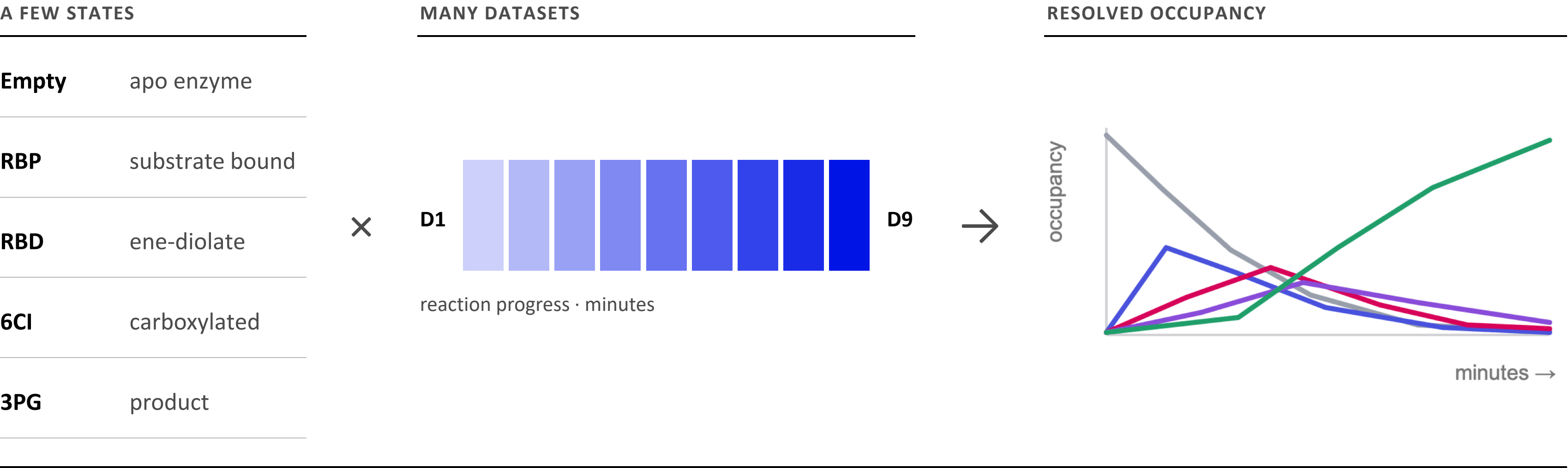


Fig. 21 — azo-CA4 (*cis*→*trans*) in $\alpha\beta$ -tubulin, 7YYZ at 10 μ s. The same ligand-unbinding system from earlier in this talk.

From difference to kinetic refinement — what RuBisCO needs

Two states generalise to many. The same multi-model, multi-dataset machinery refines a handful of conformational states simultaneously against an entire time series — letting their populations vary.



Acknowledgements

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Philip Johnson

PSI Center for Life Sciences

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Most of the RuBisCO work shown today is Georgii Khusainov’s.

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Natacha Gaillard

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Universität Frankfurt

Shapiro Group

TU Dortmund

MFX Team · LCLS

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